



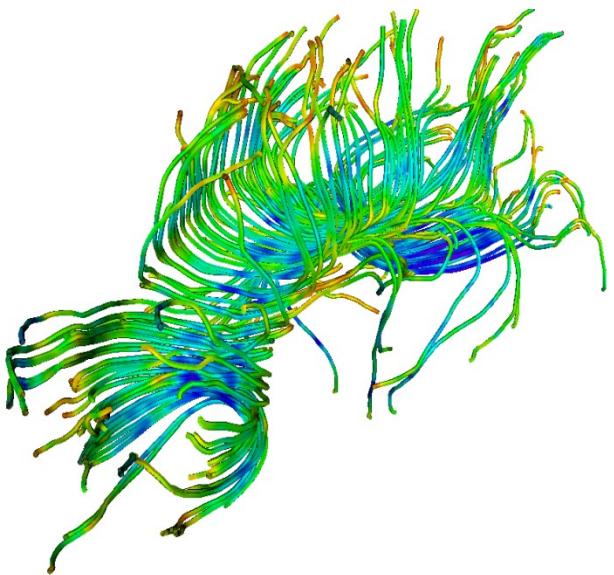
NA-MIC

National Alliance for Medical Image Computing

<http://na-mic.org>

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# Diffusion Tensor Imaging tutorial



Sonia Pujol, PhD

Surgical Planning Laboratory  
Harvard University

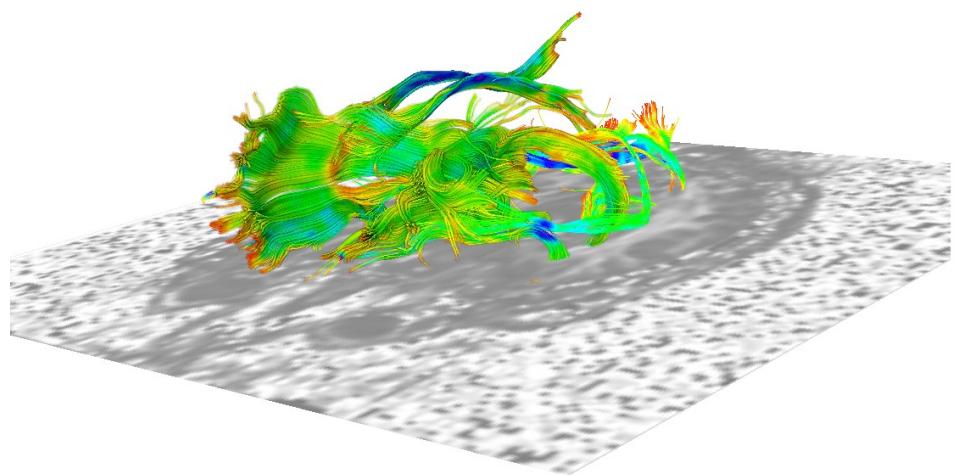
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# DTI tutorial

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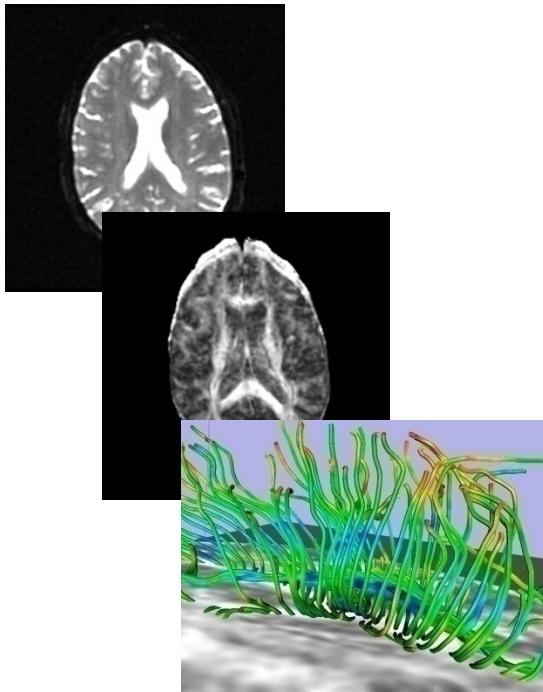
This tutorial is an introduction to the advanced **Diffusion MR** capabilities of the **Slicer3** software for medical image analysis.





# Outline

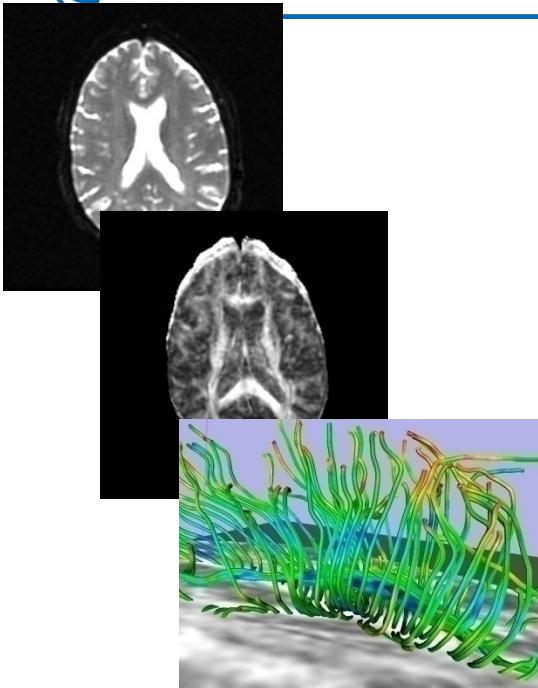
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This tutorial guides you through the process of **loading diffusion MR data**, **estimating diffusion tensors**, and **performing tractography** of white matter bundles.



# Outline



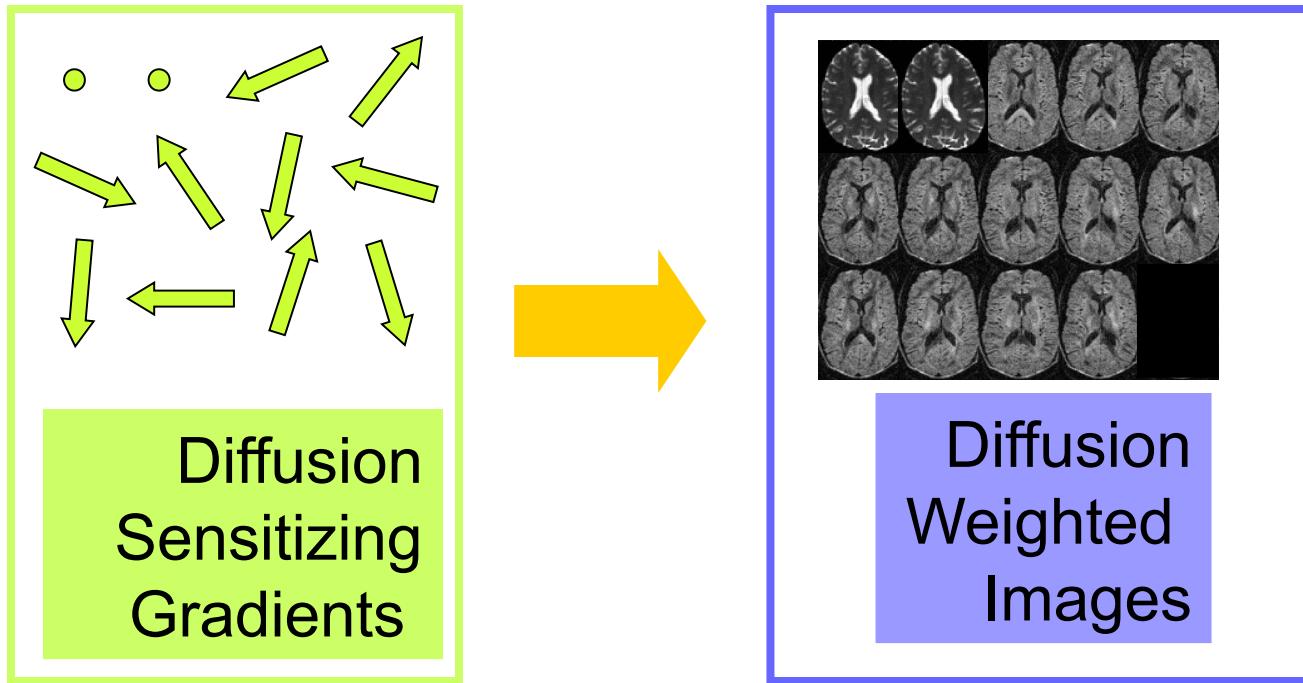
The processing pipeline uses 9 image analysis modules of Slicer3.6

1. Data
2. Volumes
3. Diffusion Tensor Estimation
4. Diffusion Tensor Scalar Measurements
5. Editor
6. LabelMap Seeding
7. Fiber Bundles
8. Fiducials
9. Fiducial Seeding



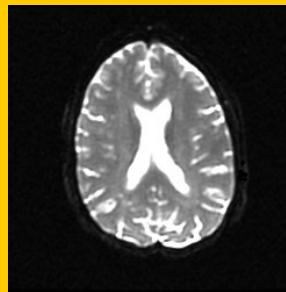
# Tutorial Dataset

The Diffusion MR tutorial dataset is composed of a **Diffusion Weighted MR scan** of the brain acquired with 12 gradient directions and 2 baseline.

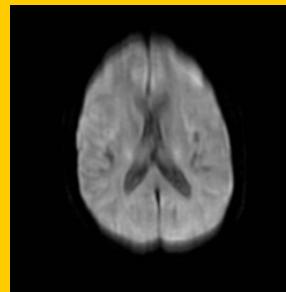




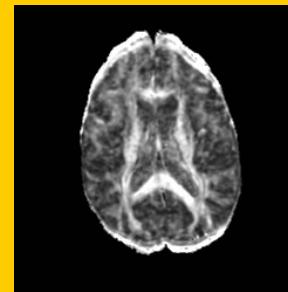
# DTI Processing Pipeline



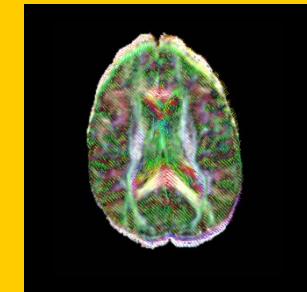
DWI  
Acquisition



Tensor  
Calculation



Scalar  
Maps



3D  
Visualization



# Start Slicer3

## Linux/Mac users

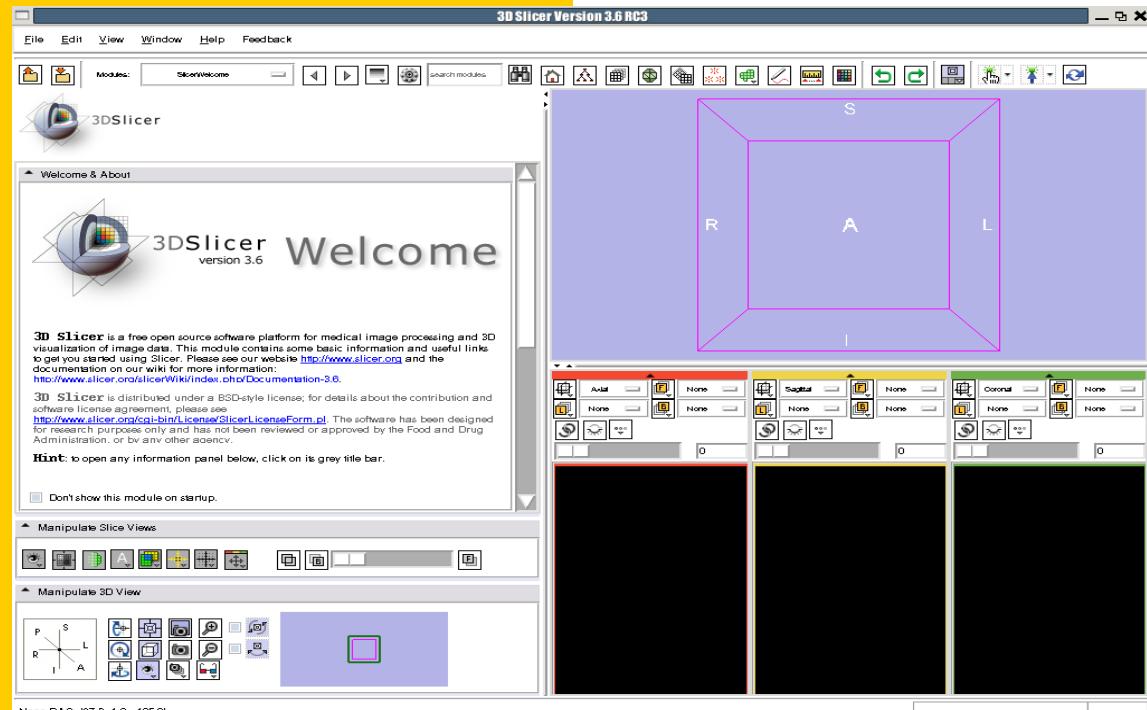
Launch the Slicer3 executable located in the Slicer3.6 directory

## Windows users

Select

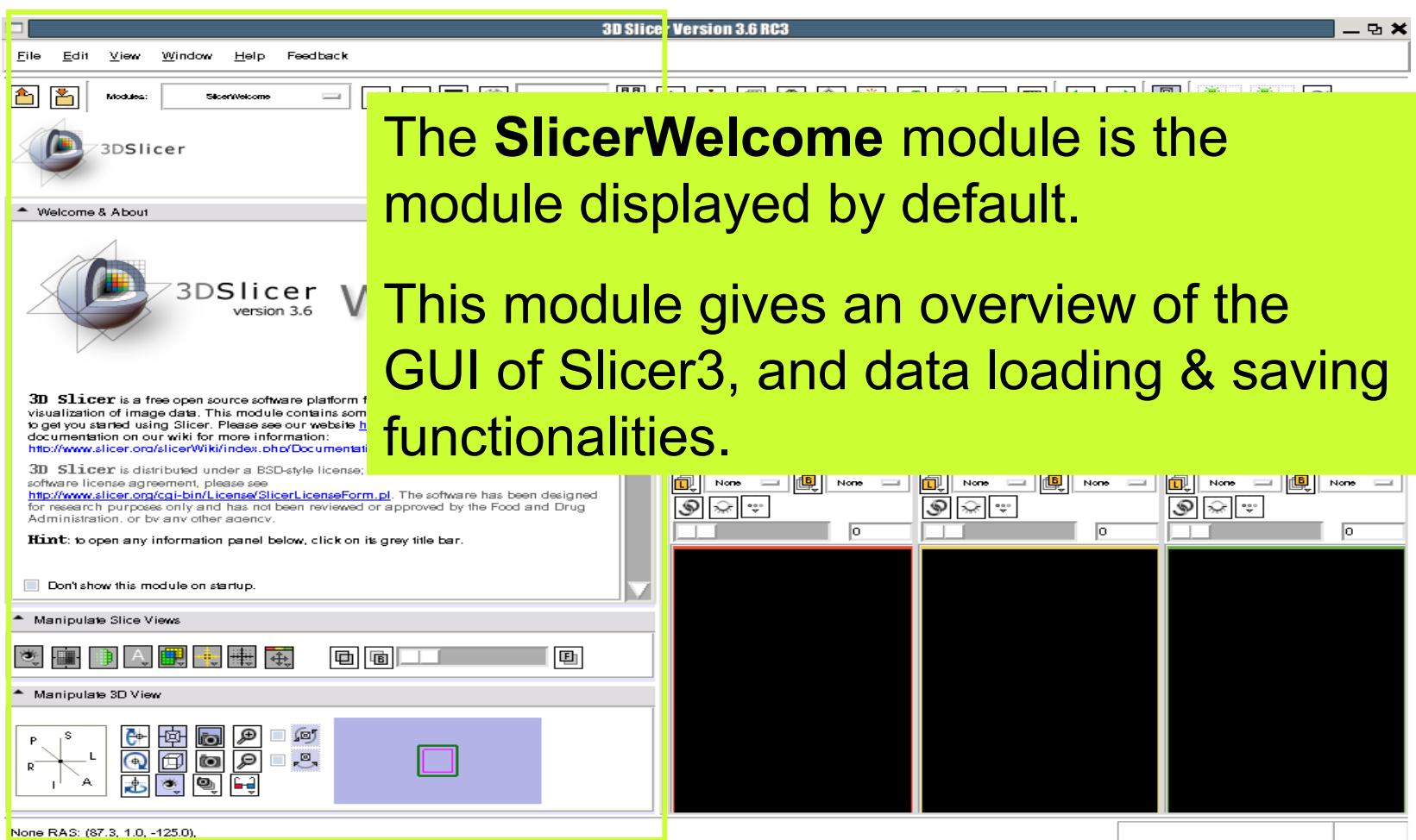
Start → All Programs

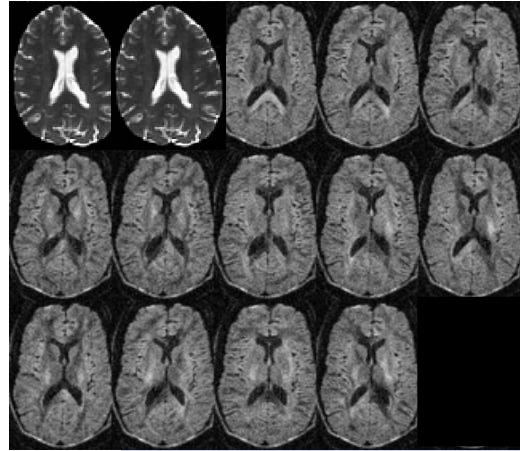
→ Slicer3-3.6-RC3-2010-06-04 → Slicer3





# Slicer Welcome



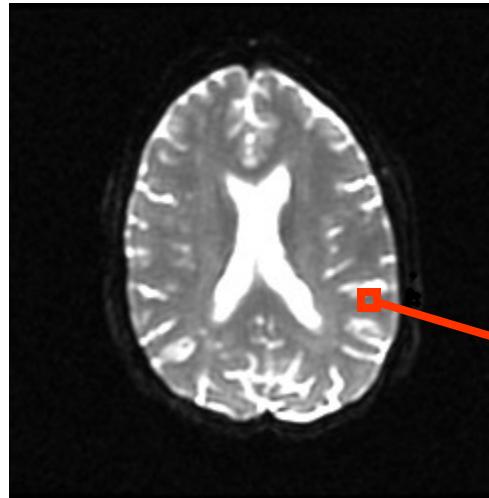


# Part 1:

# Diffusion data loading and tensor estimation



# Diffusion Tensor



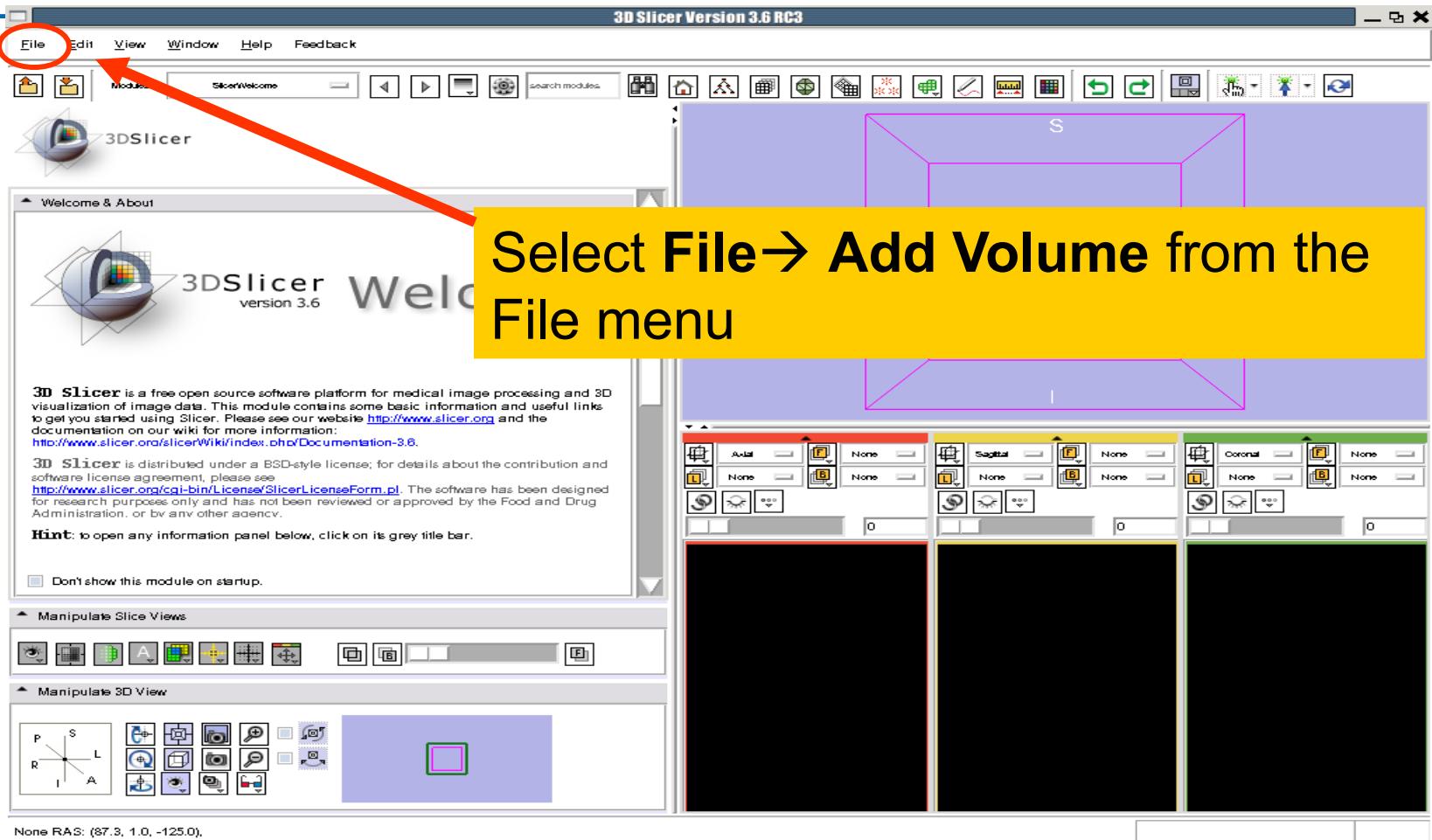
Stejskal-Tanner

$$S_i = S_0 e^{-b \hat{g} i^T D \hat{g}_i}$$

$$\underline{D} = \begin{bmatrix} D_{xx} & D_{xy} & D_{xz} \\ D_{yx} & D_{yy} & D_{yz} \\ D_{zx} & D_{zy} & D_{zz} \end{bmatrix}$$

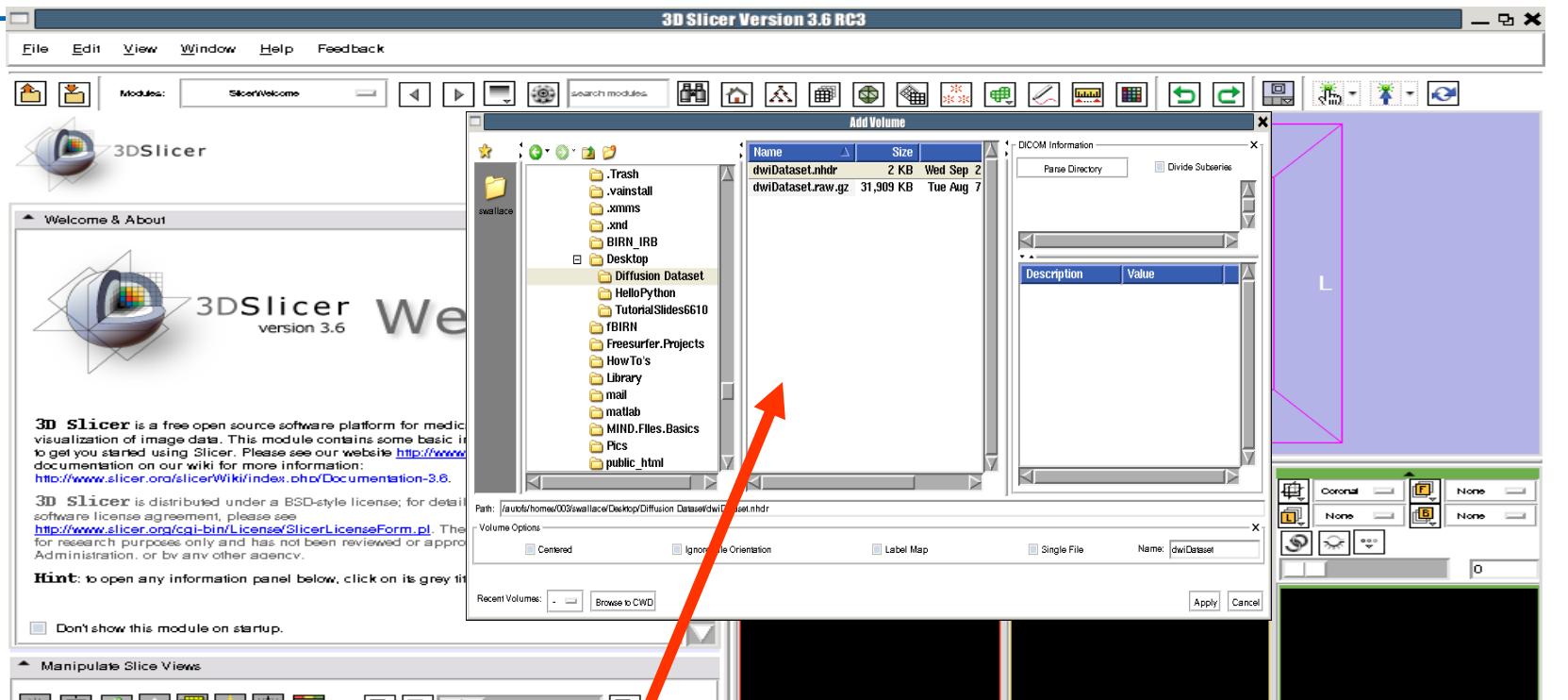


# Loading the DWI volume





# Loading the DWI volume

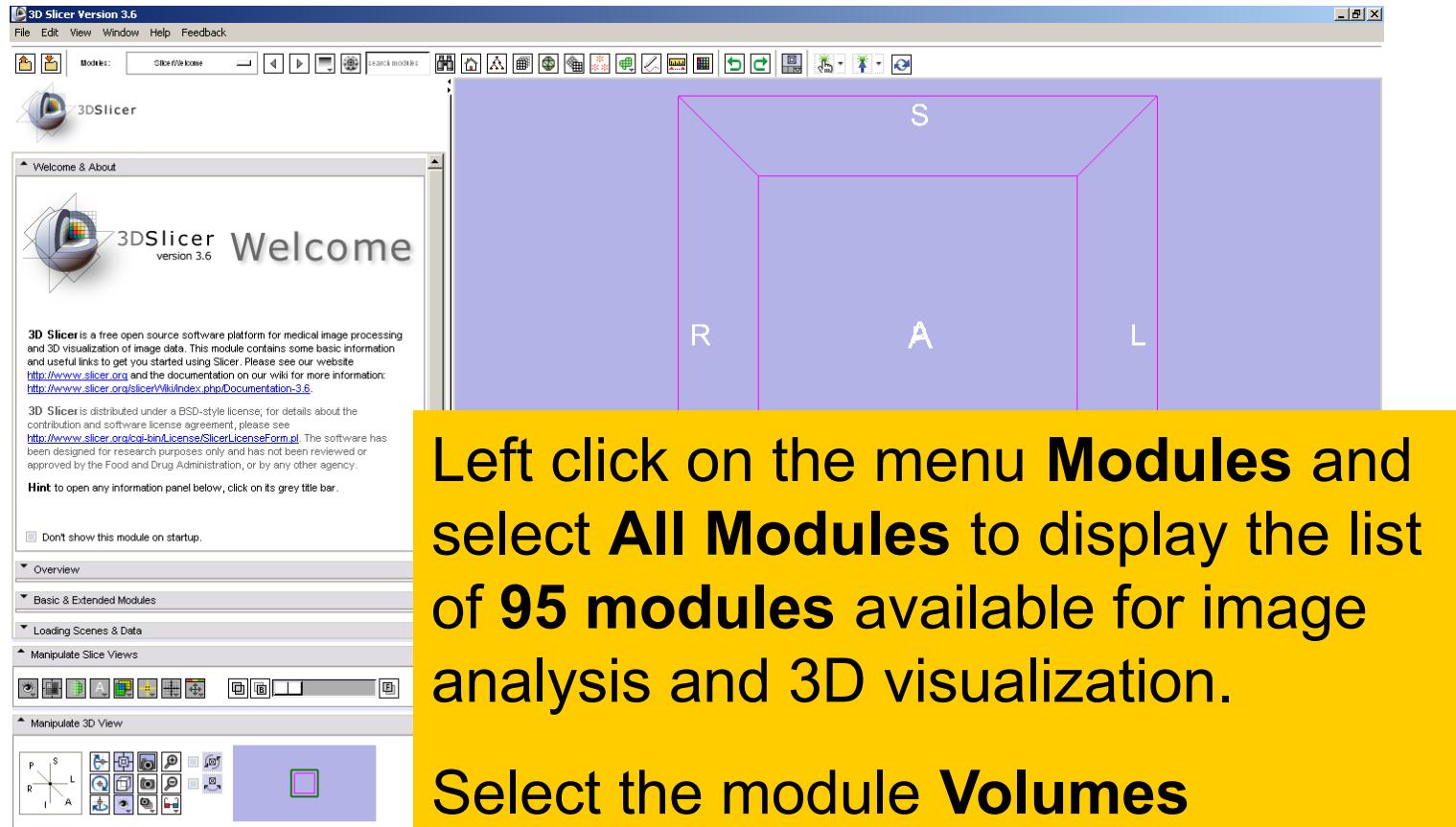


Browse to the location of the Diffusion tutorial dataset directory and select the file **dwiDataset.nhdr**

Click on **Apply** to load the volume

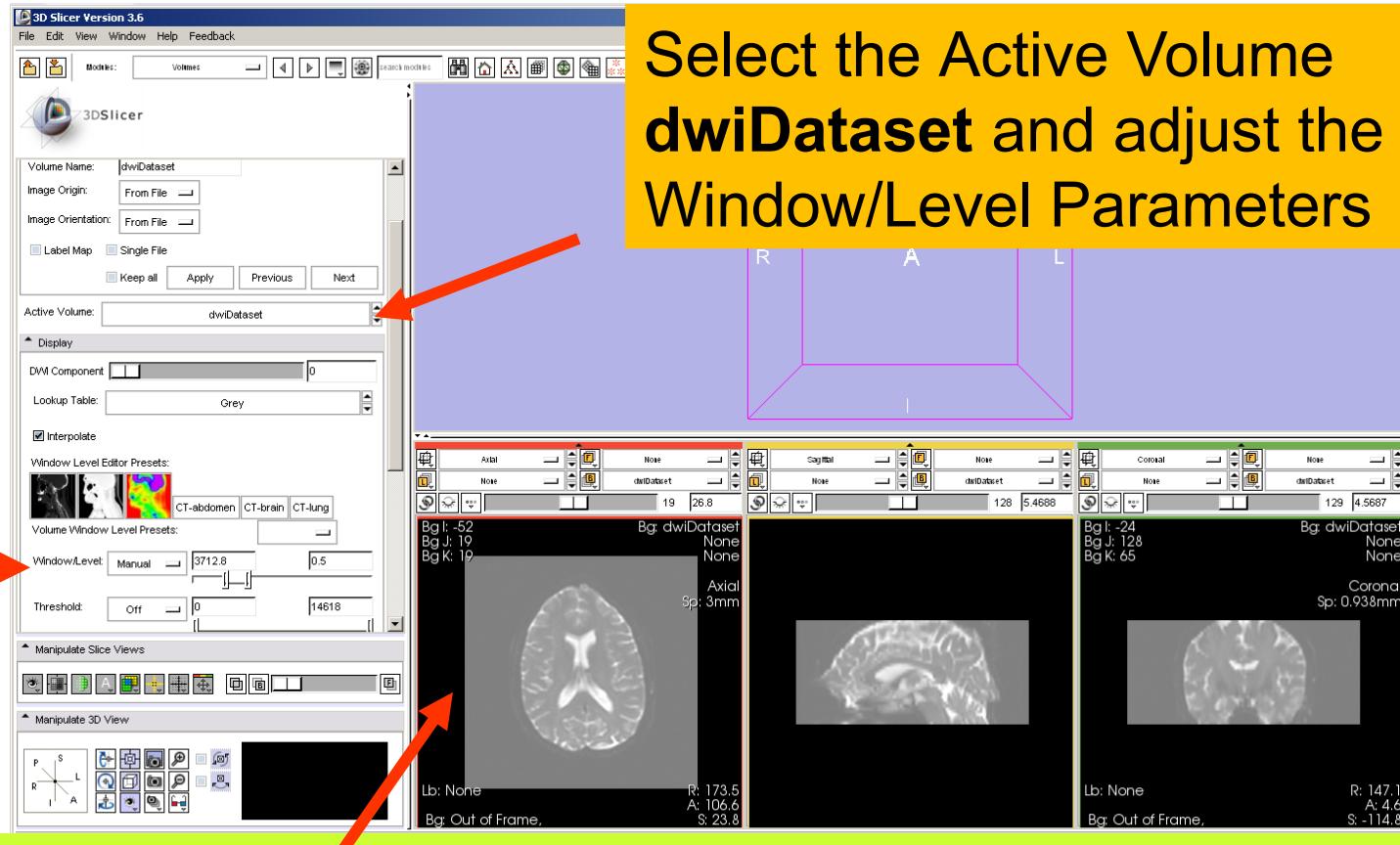


# Loading the DWI volume





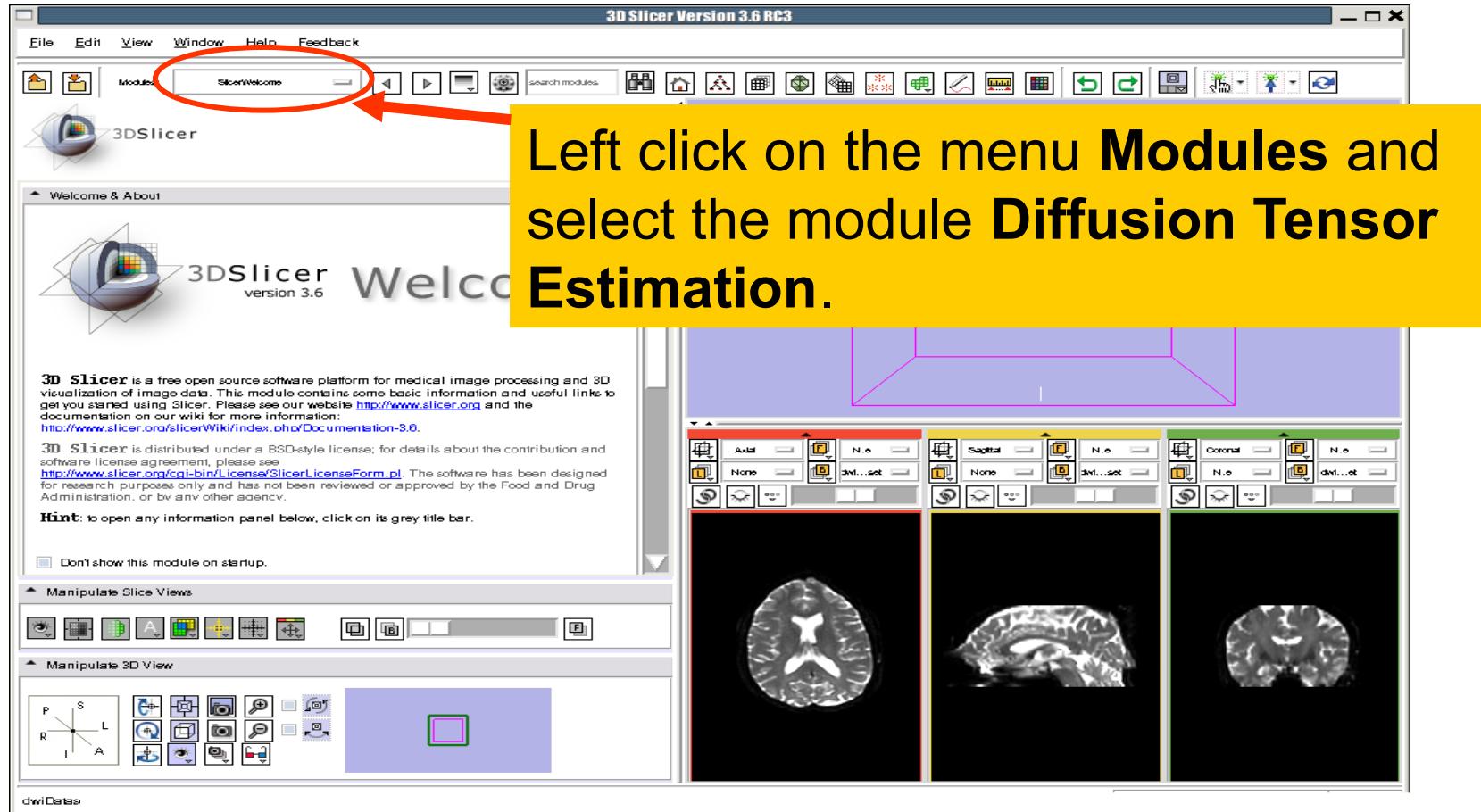
# Loading the DWI volume



Slicer displays the anatomical views of the baseline volume of the diffusion dataset in the 2D Slice Viewer.

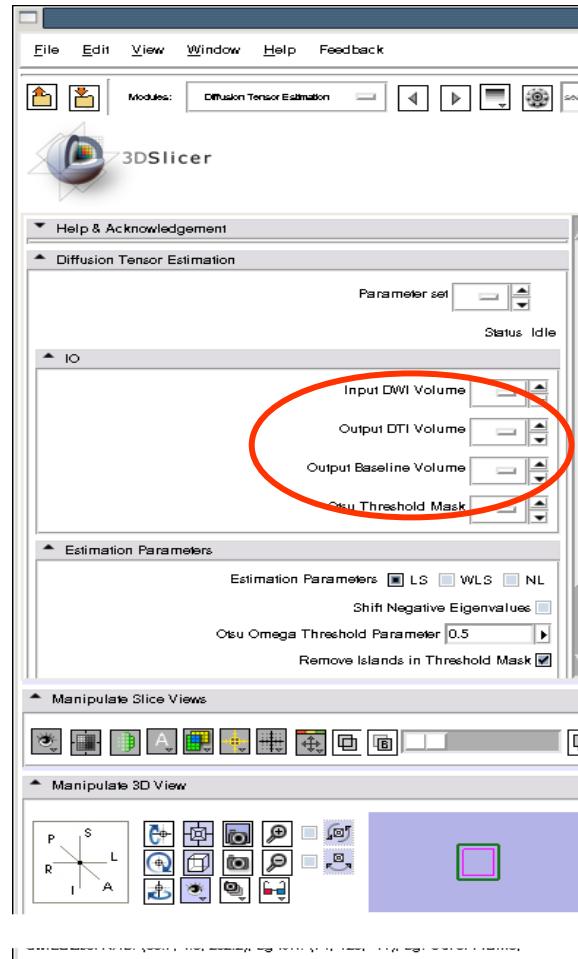


# Tensor Estimation





# Tensor Estimation



Select the Input DWI Volume  
`dwiDataset`

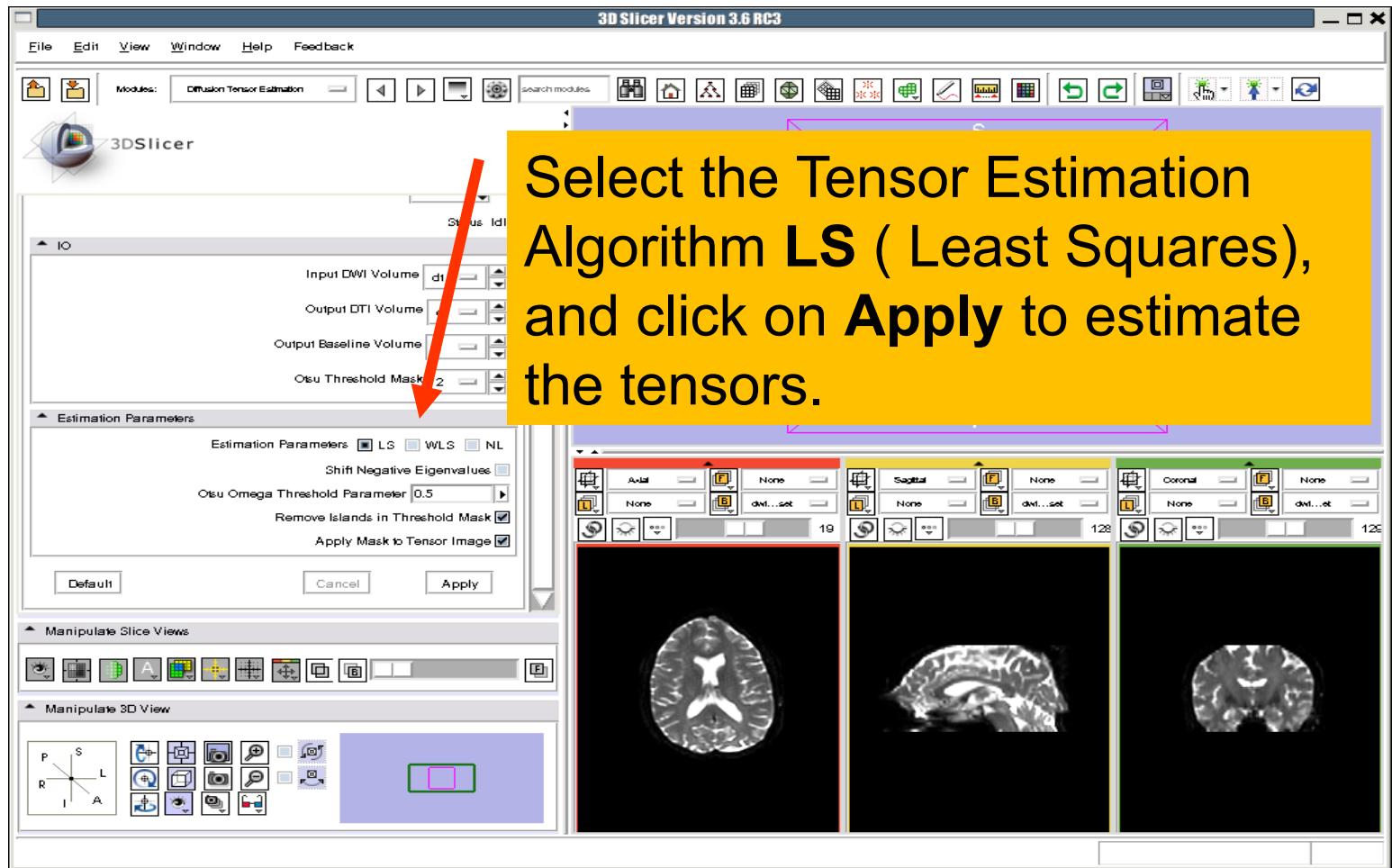
Left click on **OutputDTIVolume** and  
select ‘Create New Diffusion  
Tensor Volume’

Left click on **Output Baseline  
Volume** and select ‘Create New  
Volume’

Left click on **Otsu Threshold Mask**  
and select ‘Create New Volume’

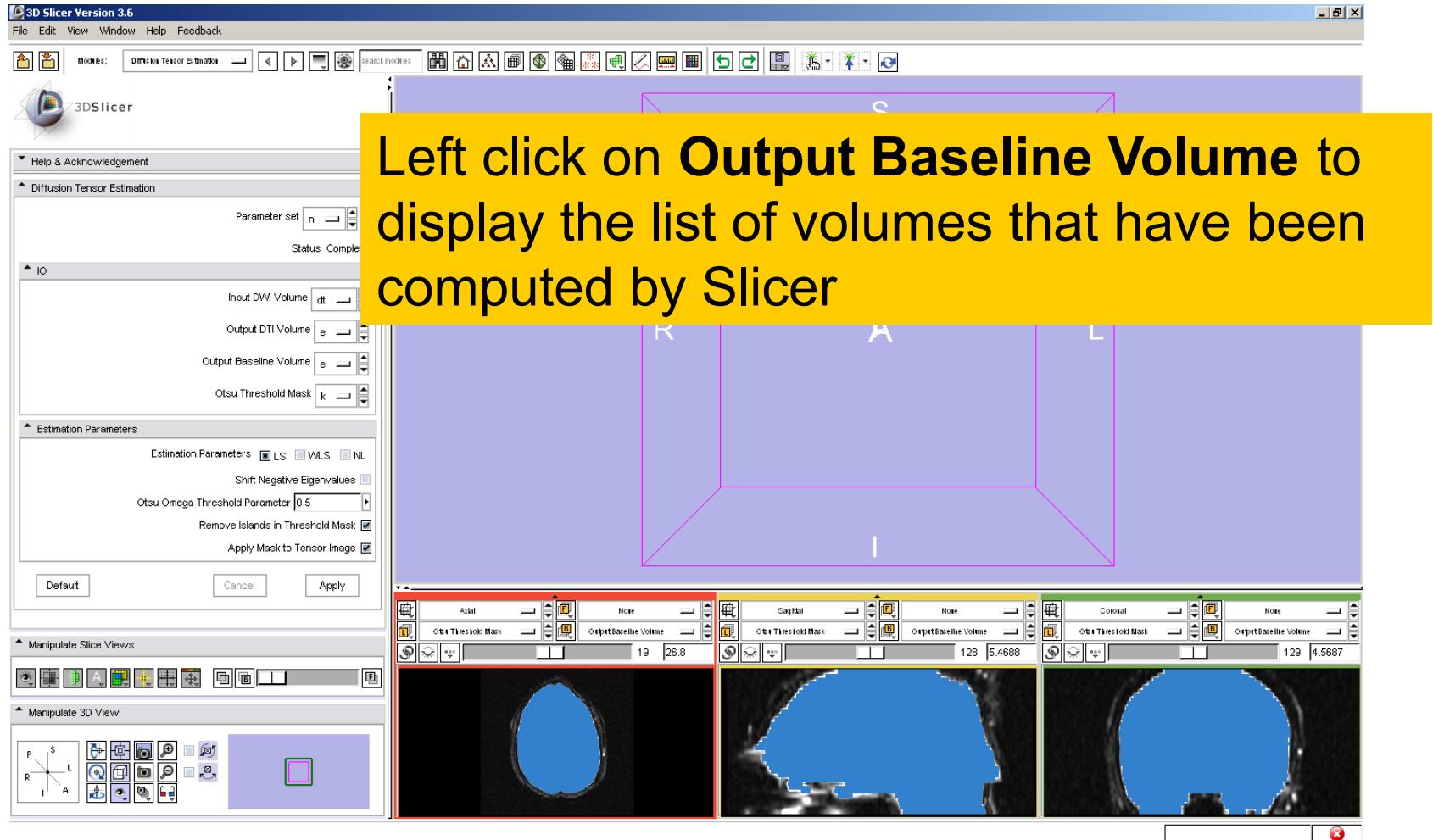


# Tensor Estimation





# Tensor Estimation



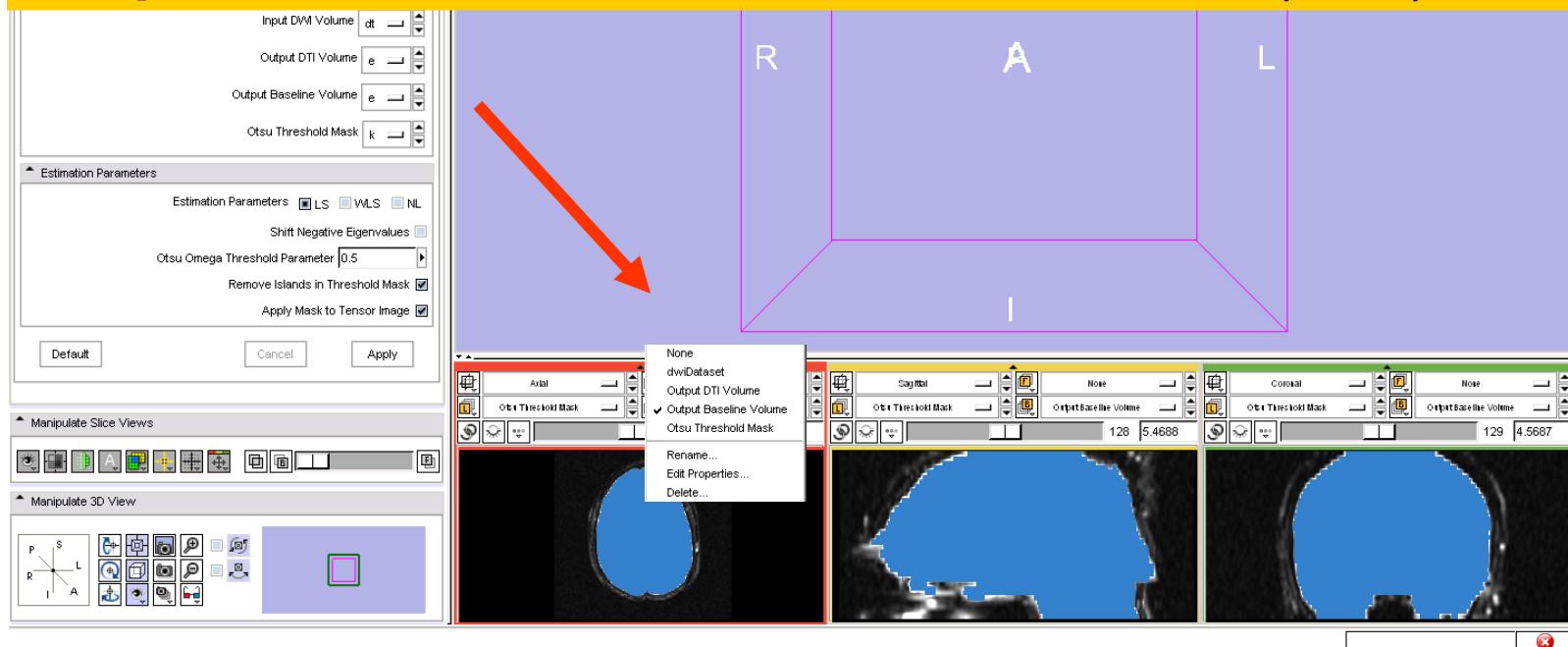


# Tensor Estimation

**Output DTI Volume** is the volume of estimated tensors

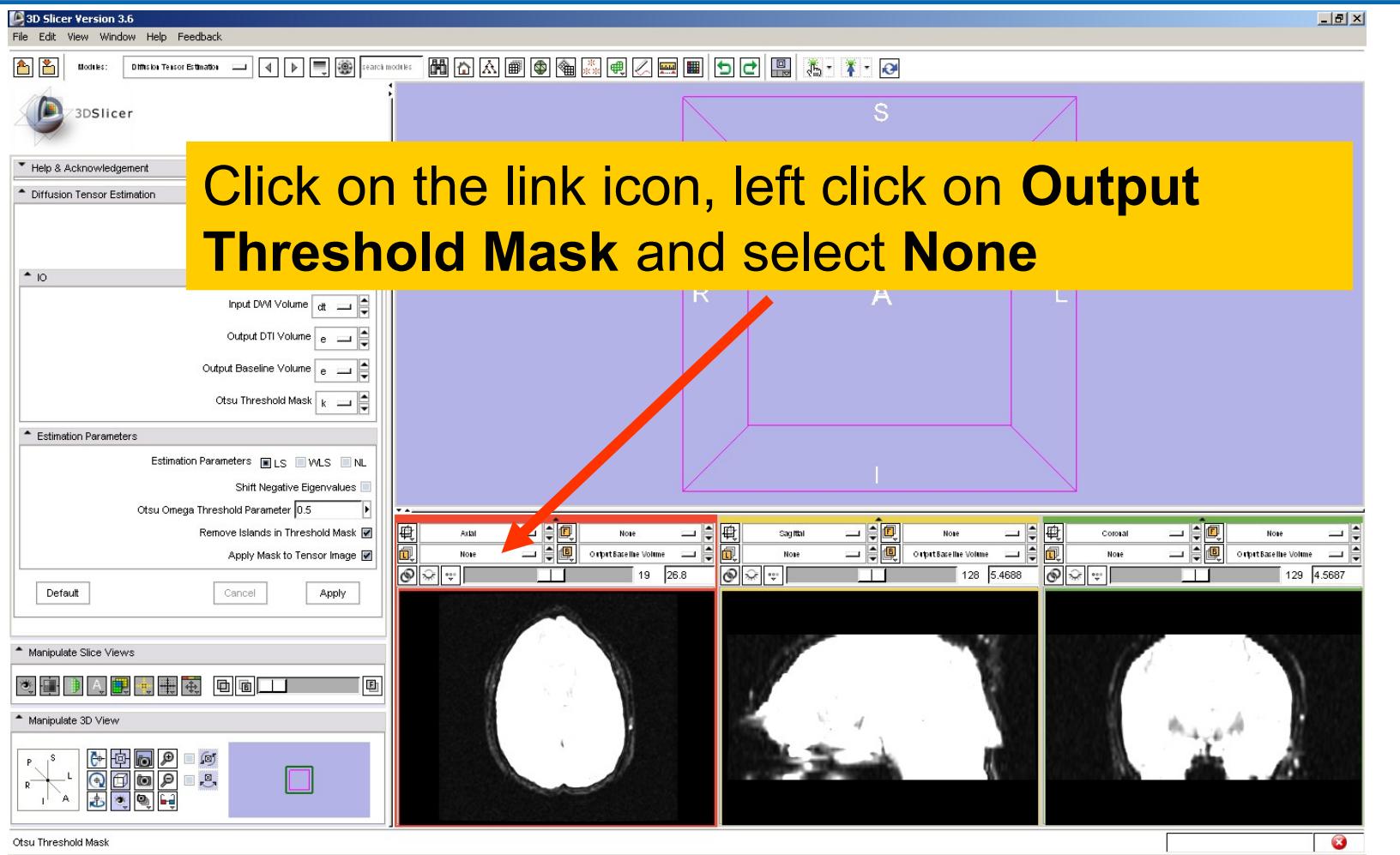
**Output Baseline Volume** is the Baseline volume

**Output Threshold Mask** is the tensor mask (blue)



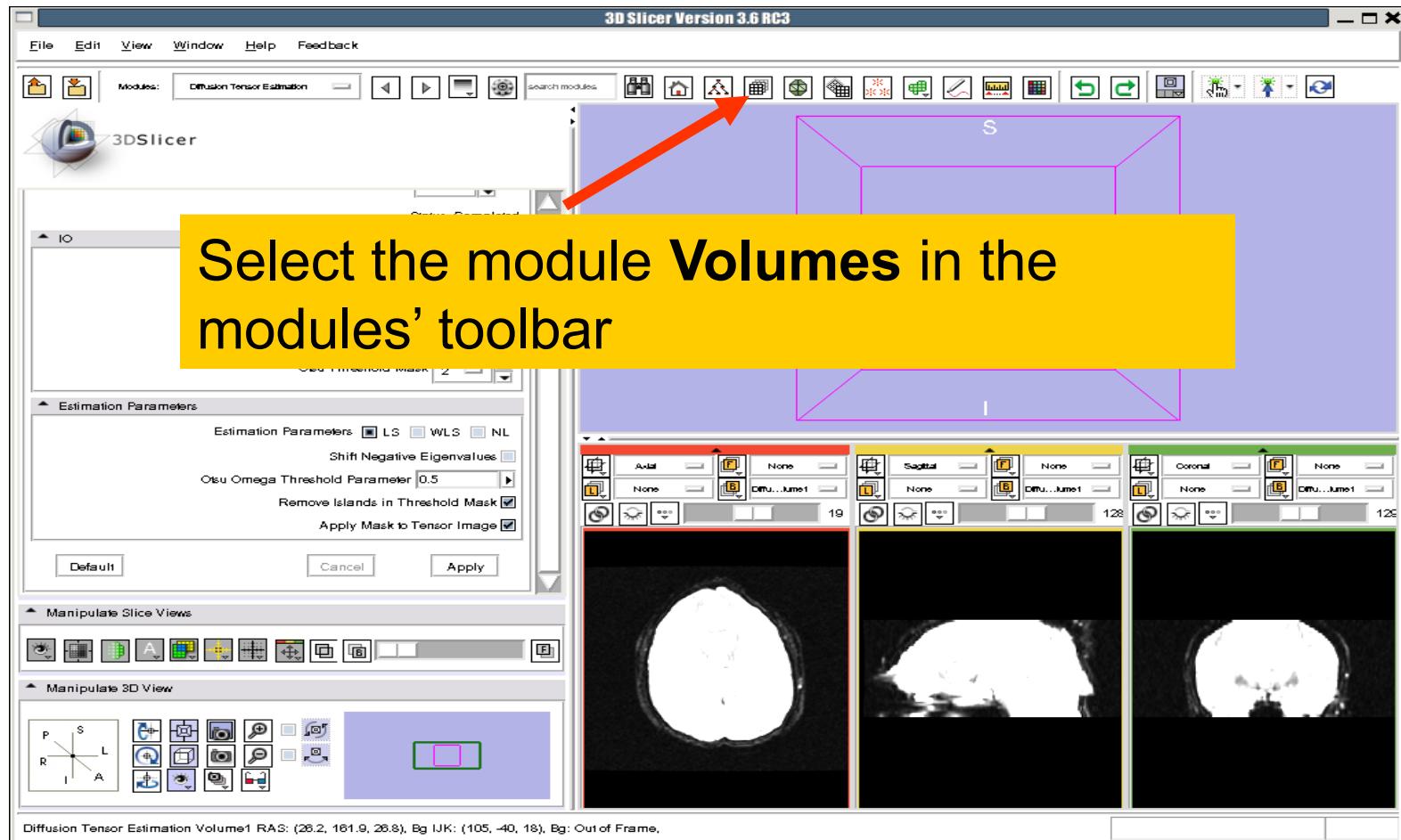


# Tensor Estimation



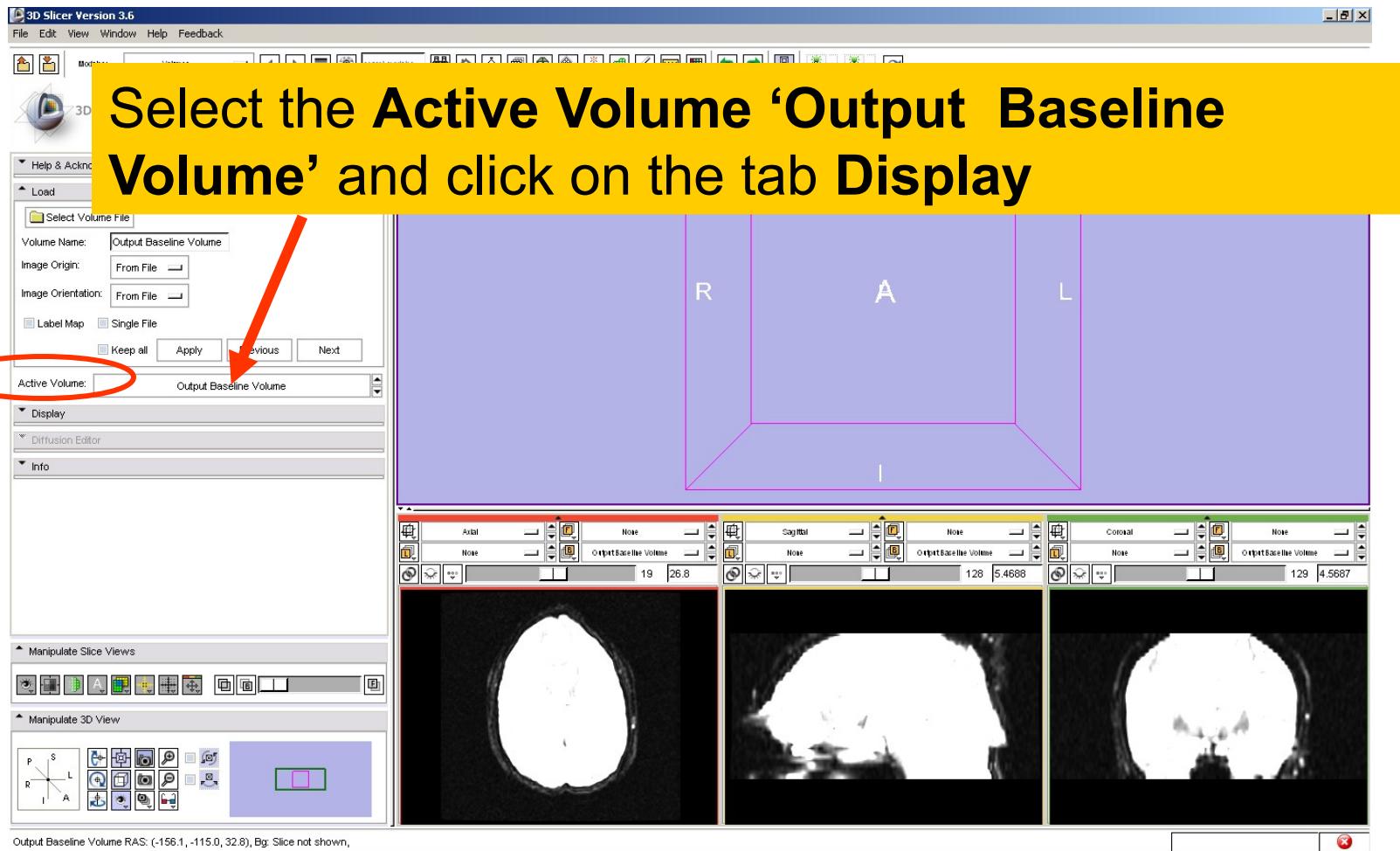


# Tensor Estimation



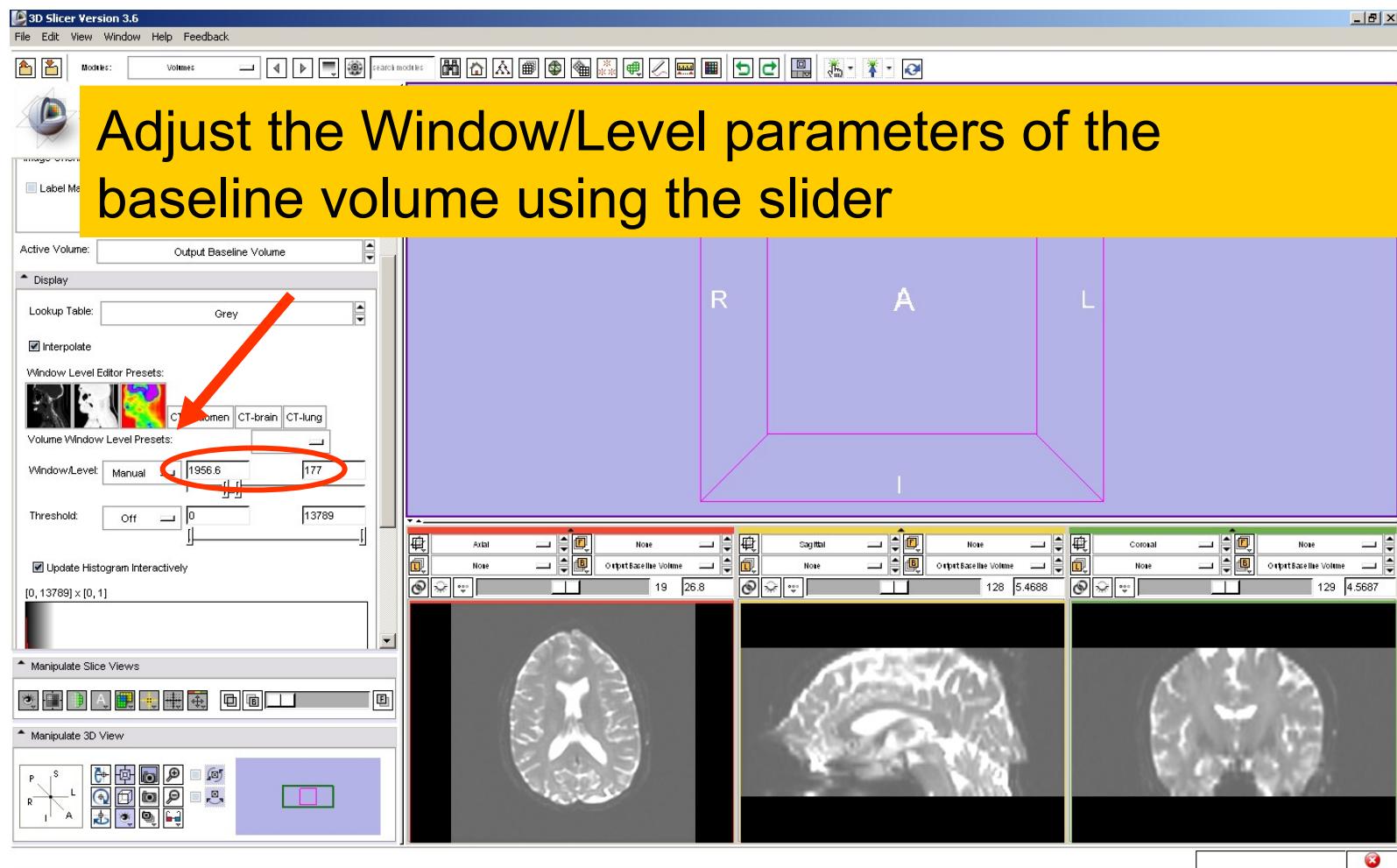


# Tensor Estimation



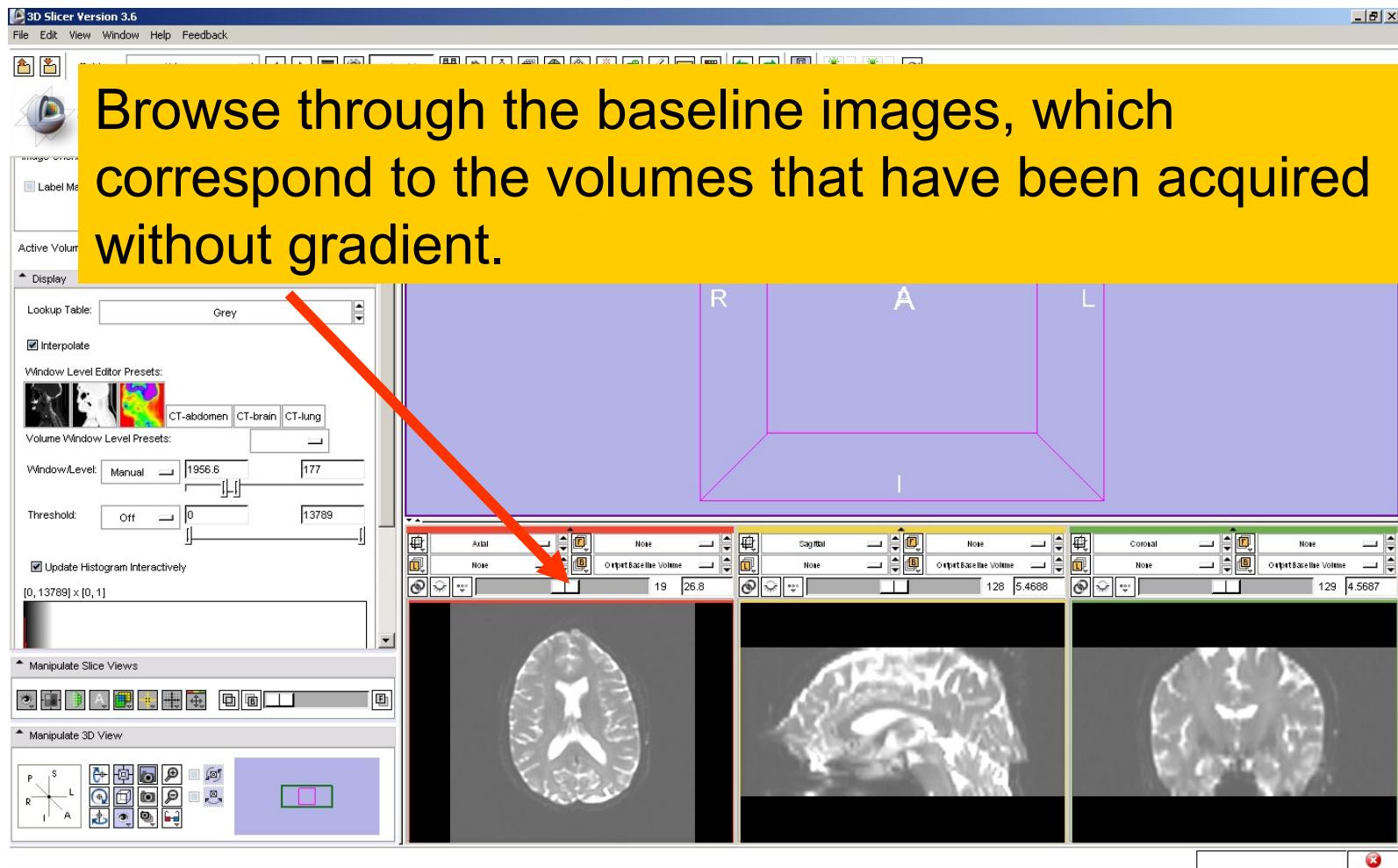


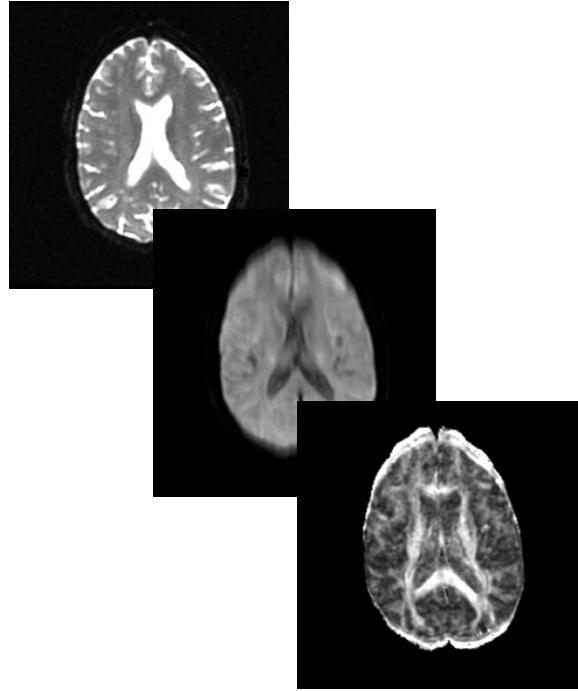
# Tensor Estimation





# Tensor Estimation





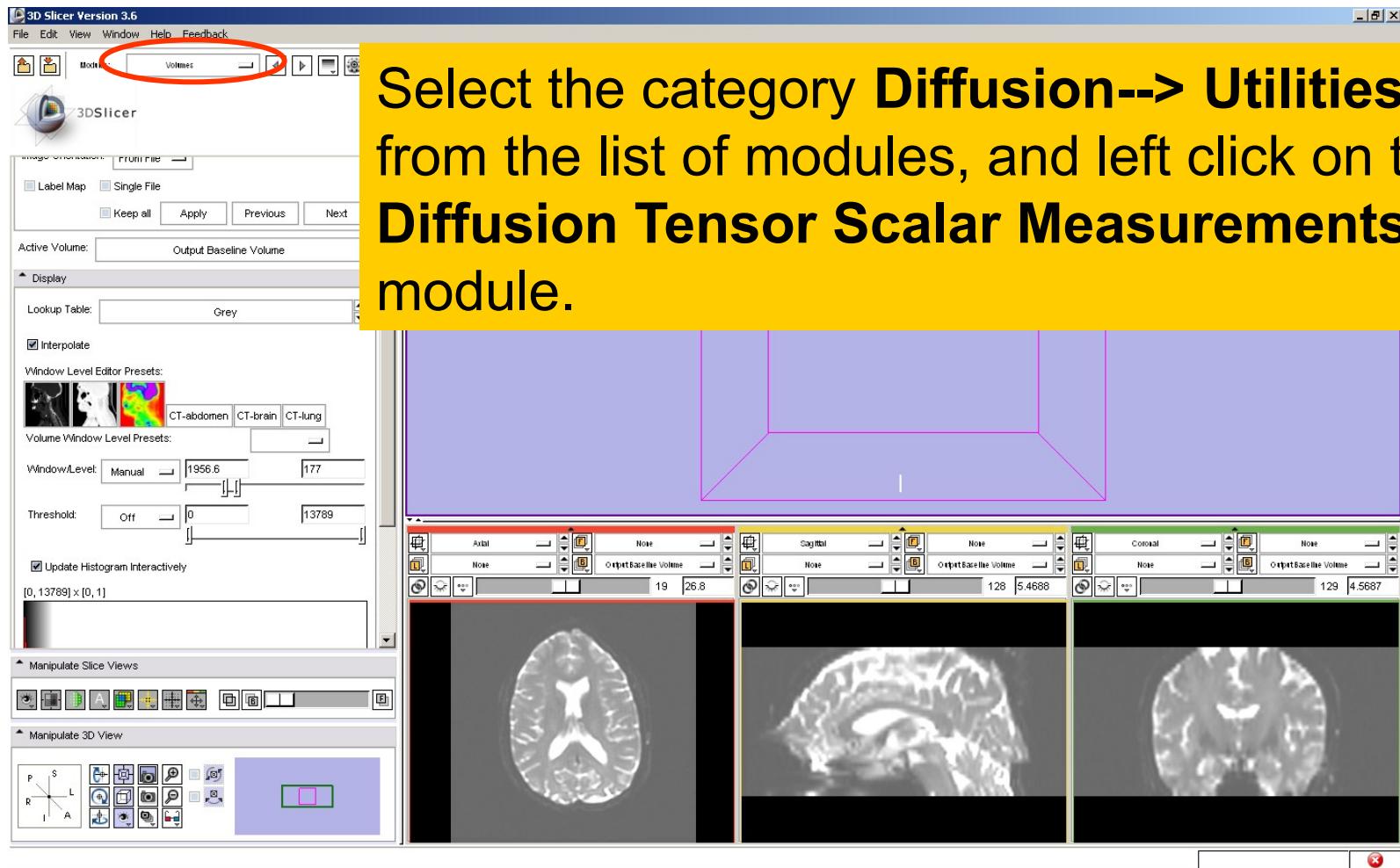
# Part2:

# Scalar

# Measurements

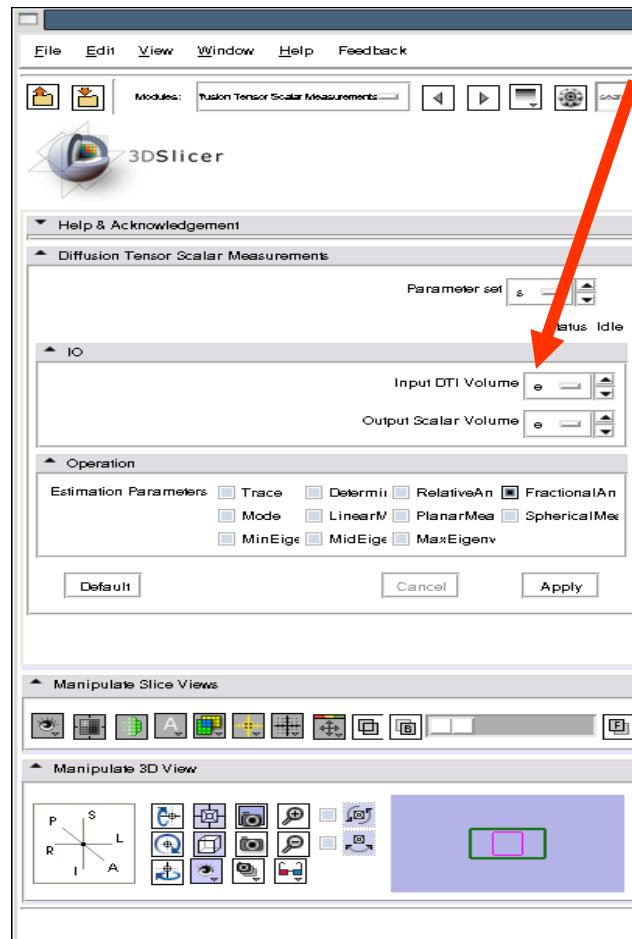


# Scalar Measurements





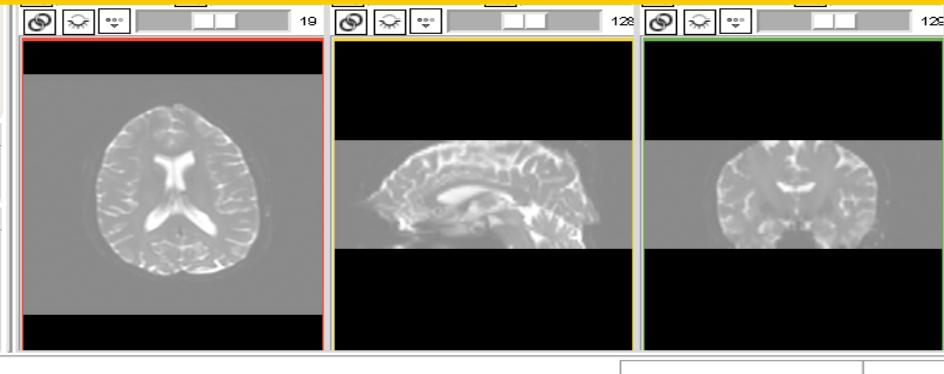
# Scalar Measurements



Select the Input DTI Volume **Output DTI Volume**

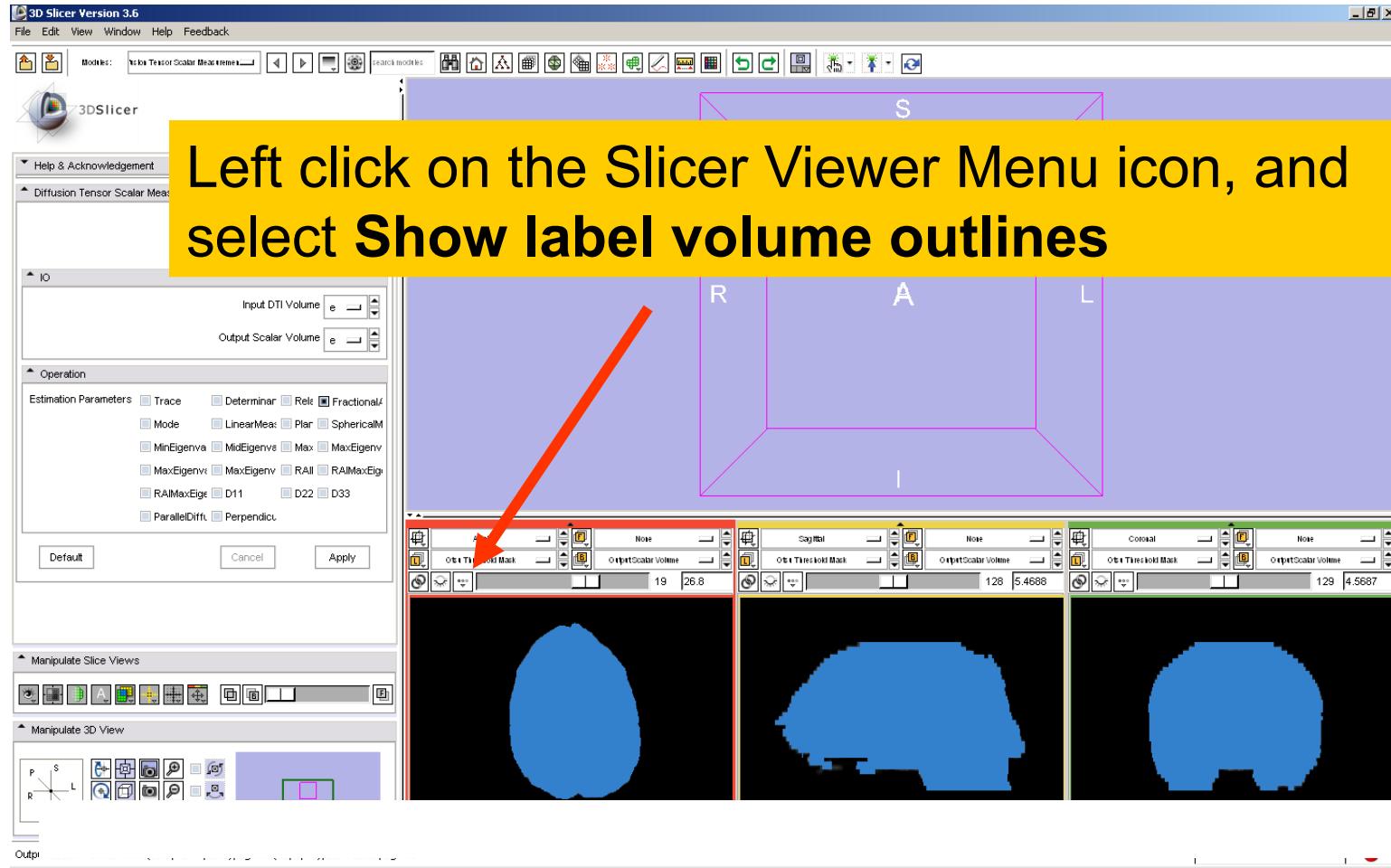
Select the Output Scalar Volume  
**'Create New Volume'**

Select the Operation **Fractional Anisotropy**, and click on **Apply**



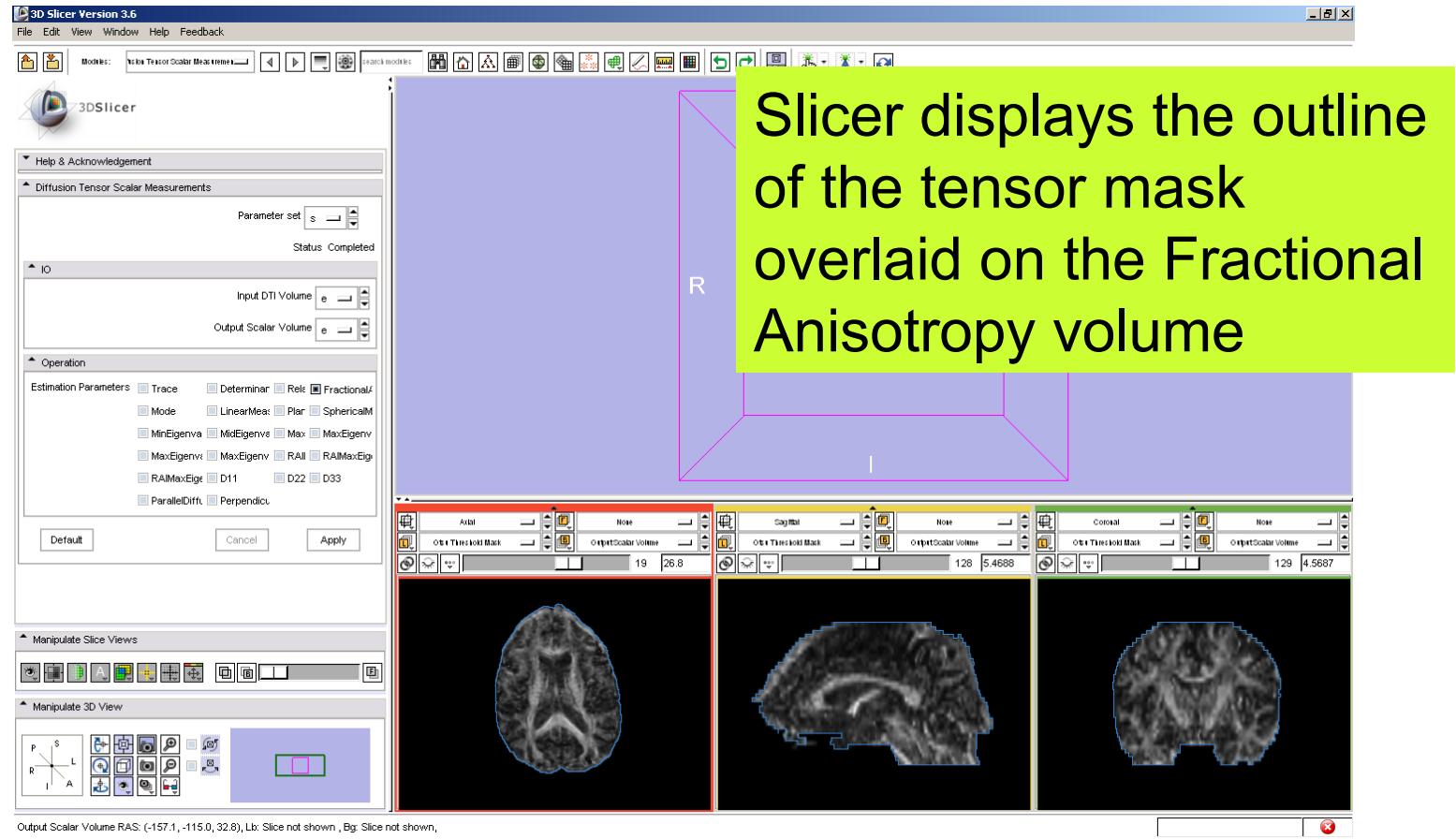


# Fractional Anisotropy Volume



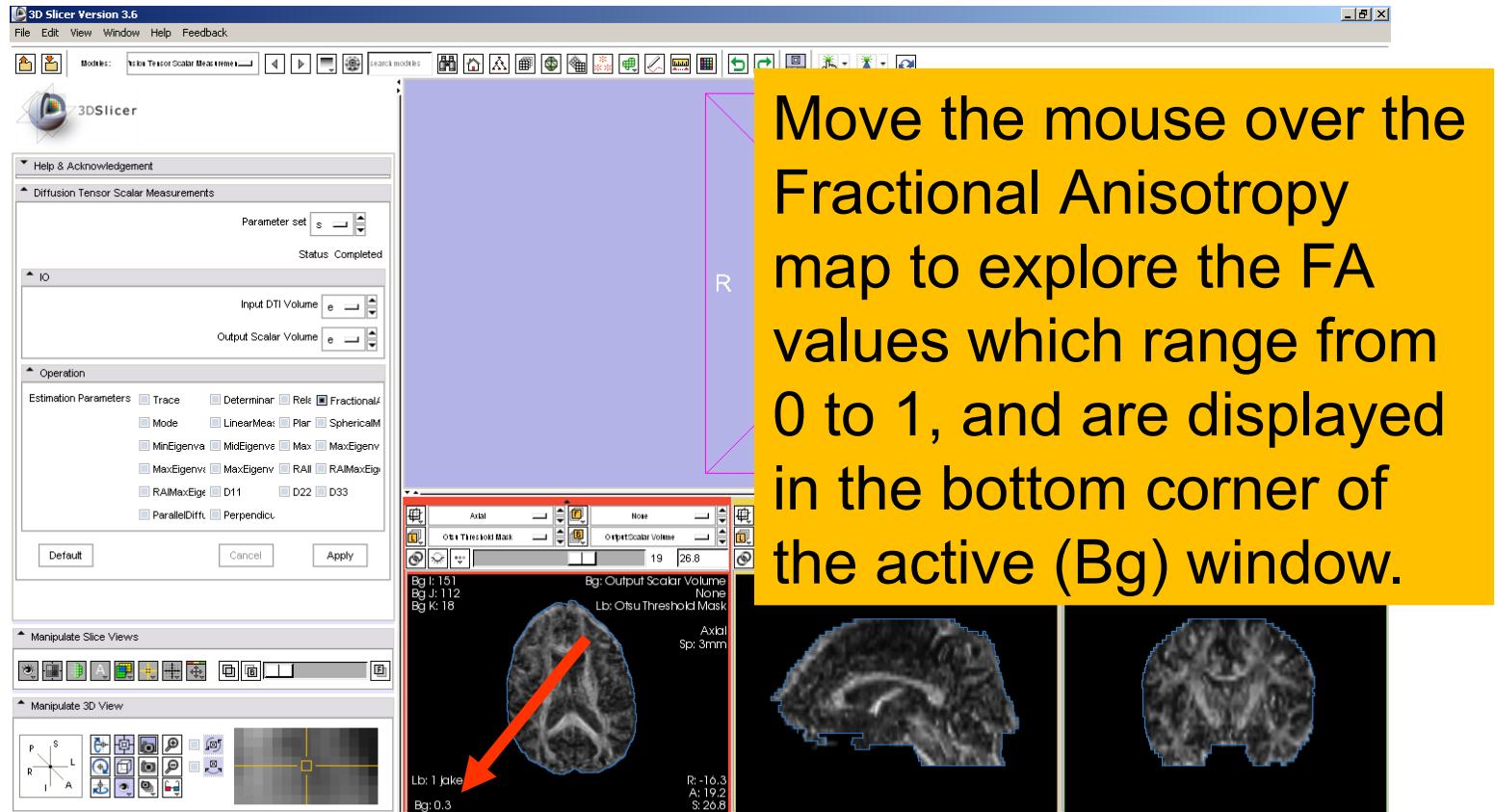


# Fractional Anisotropy Volume





# Fractional Anisotropy Volume





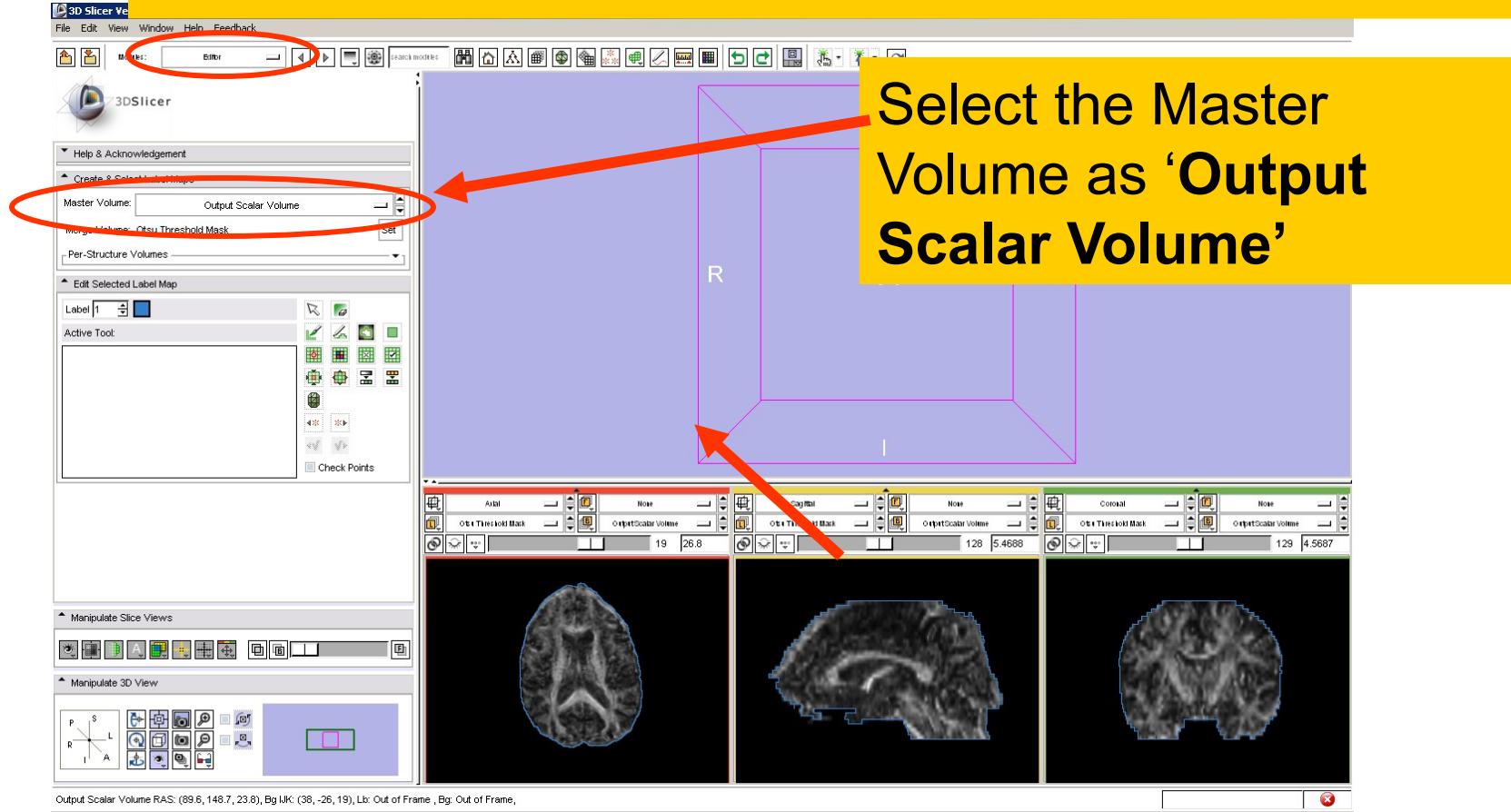
# Part 3:

# Region of Interest based Tractography



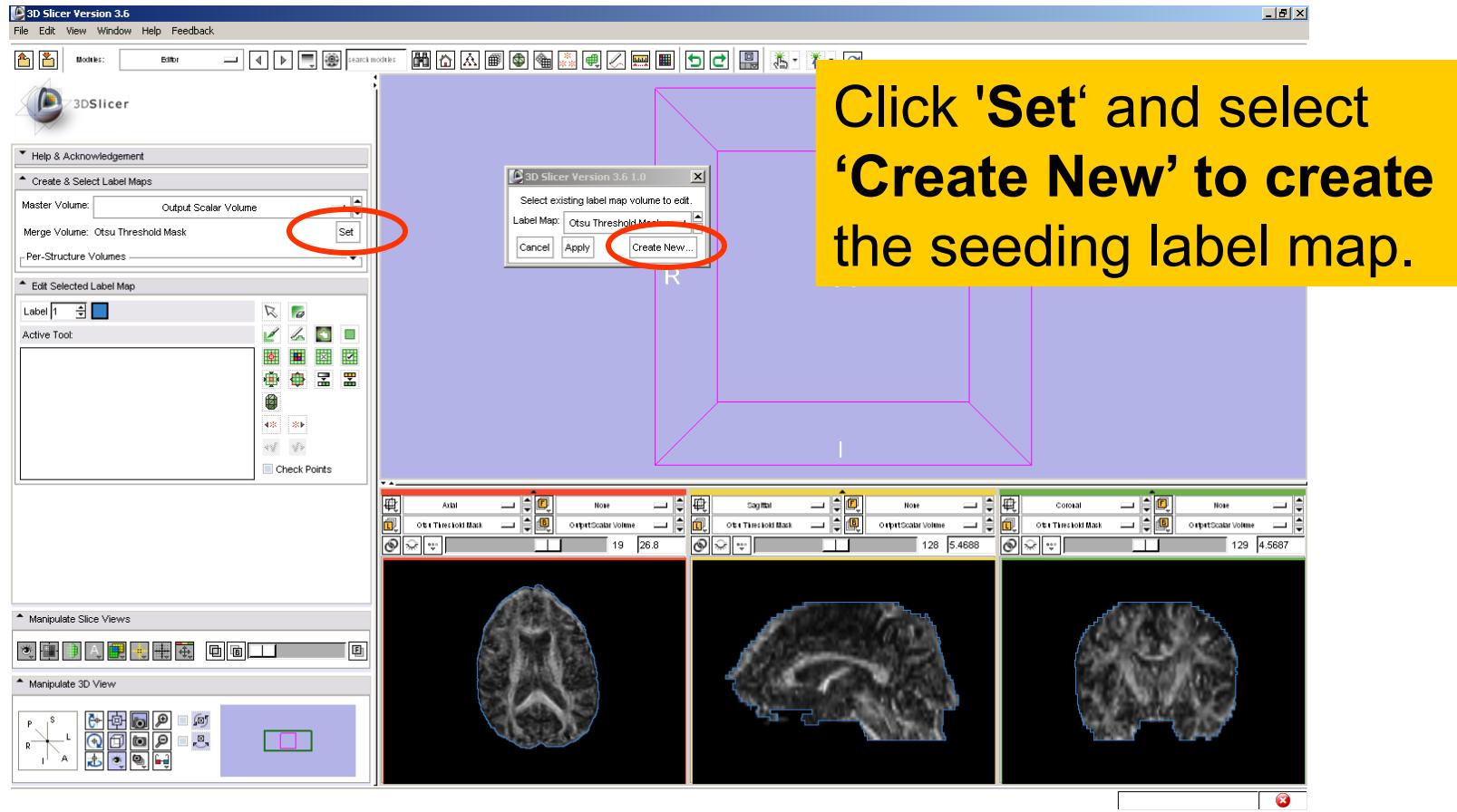
# LabelMap Generation

Select the module **Editor** in the modules menu.



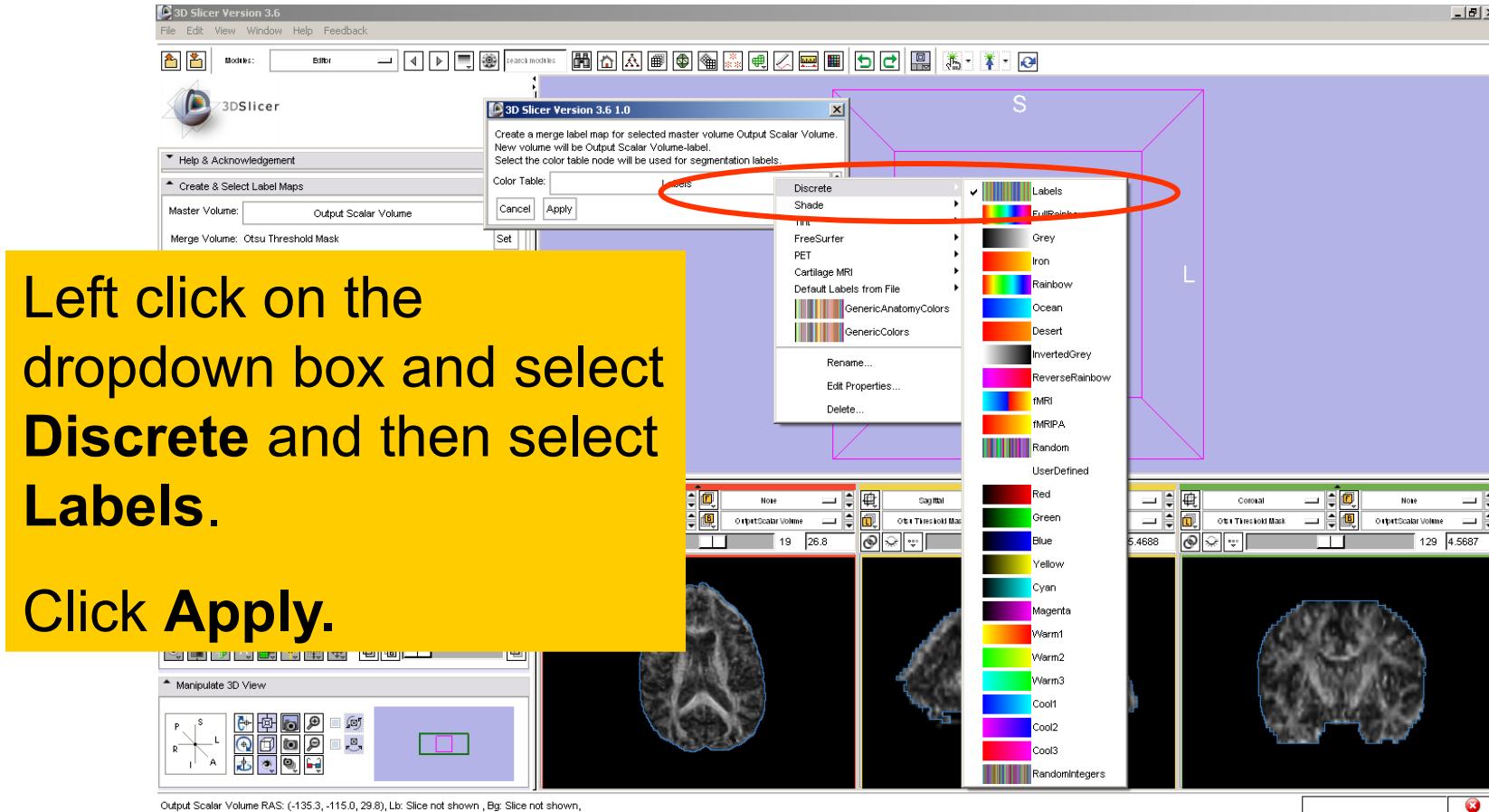


# Label Map Generation



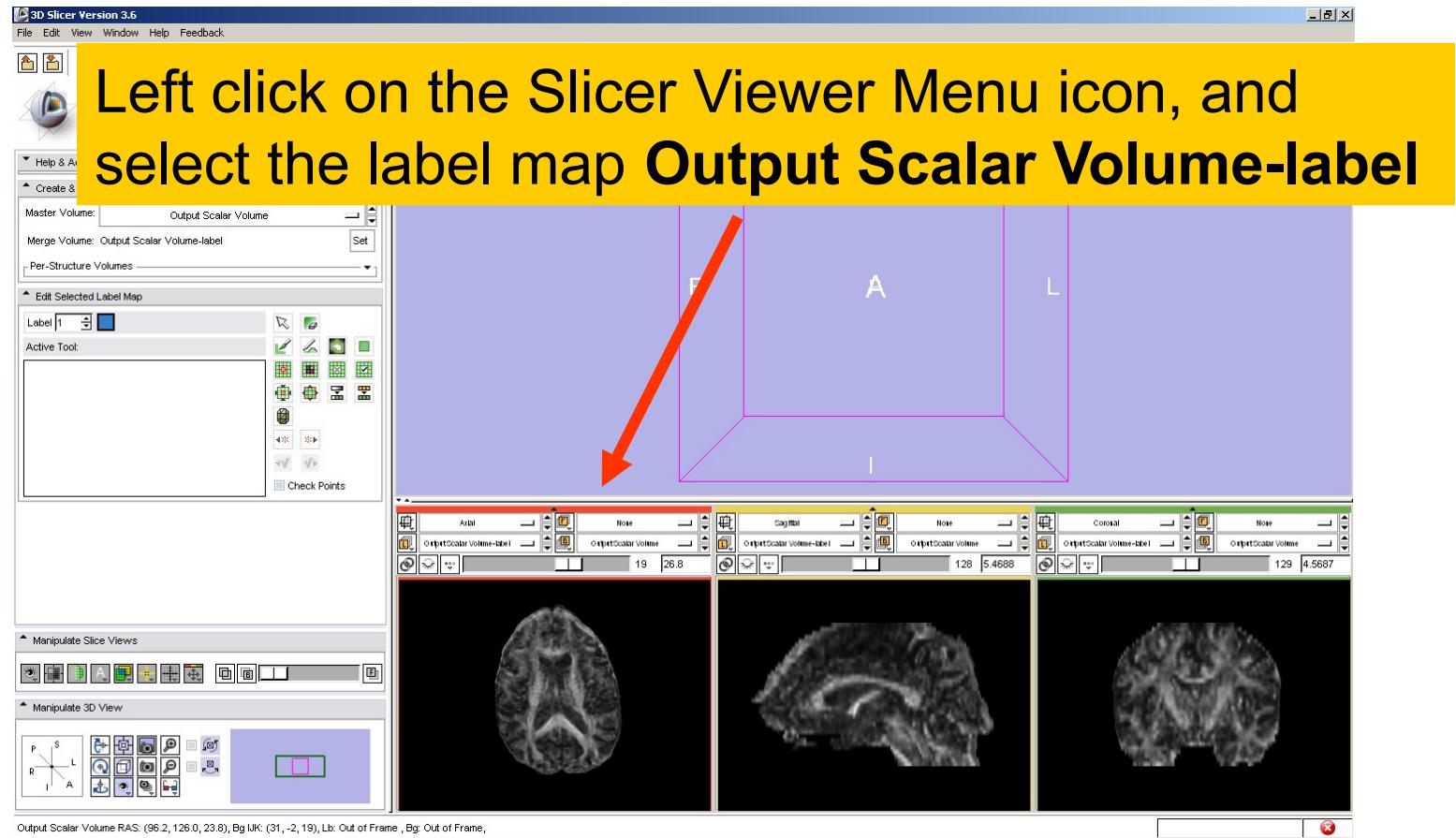


# LabelMap Generation



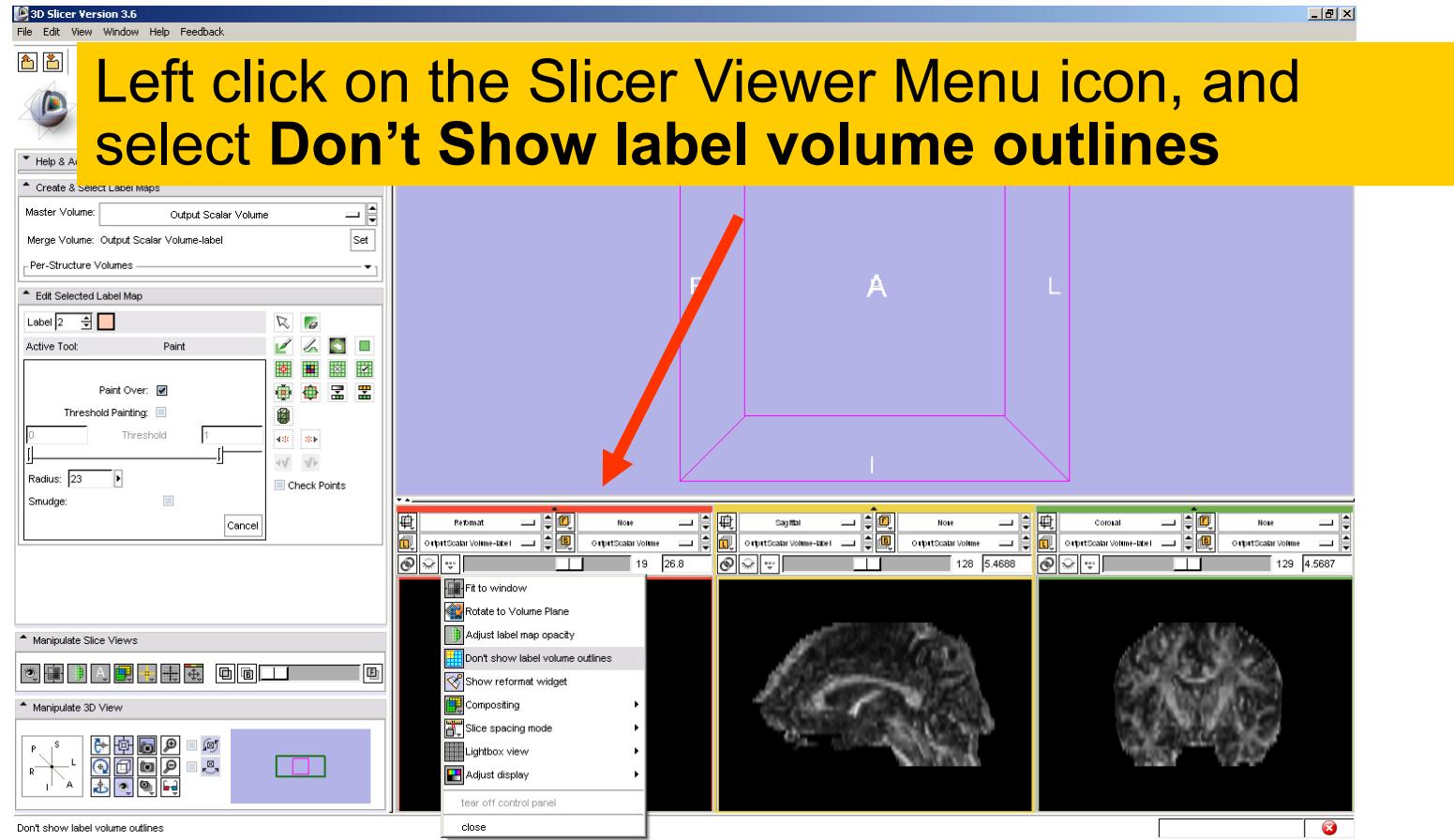


# LabelMap Generation





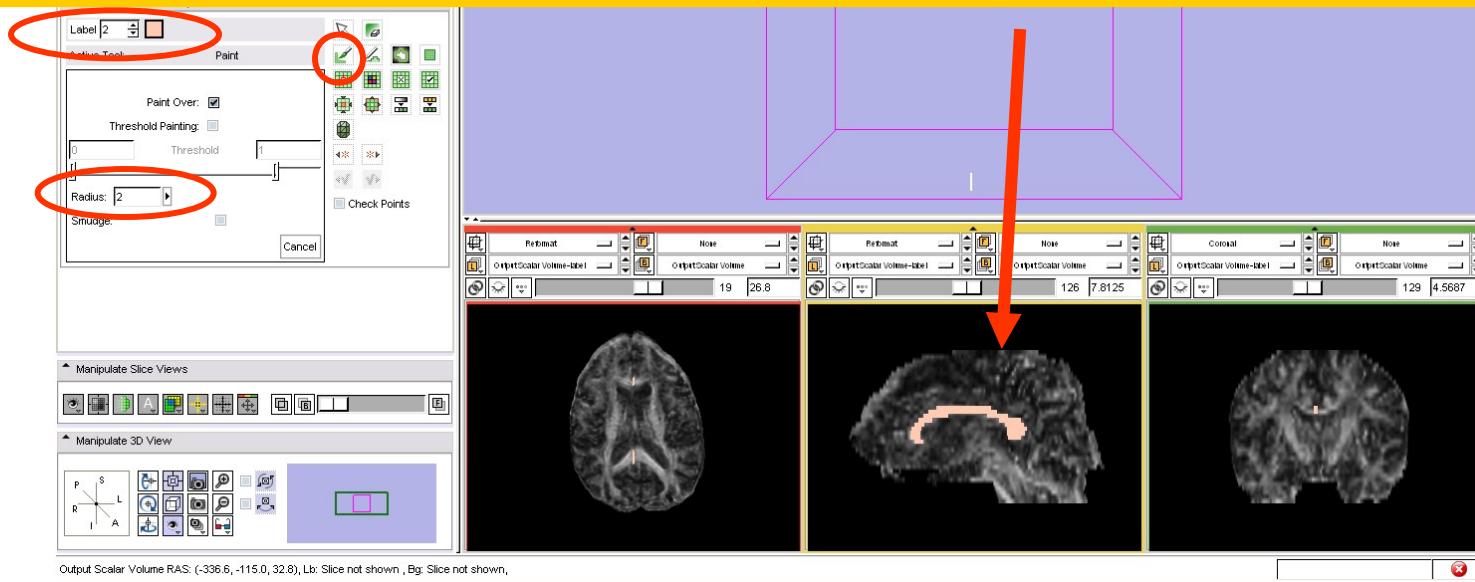
# LabelMap Generation





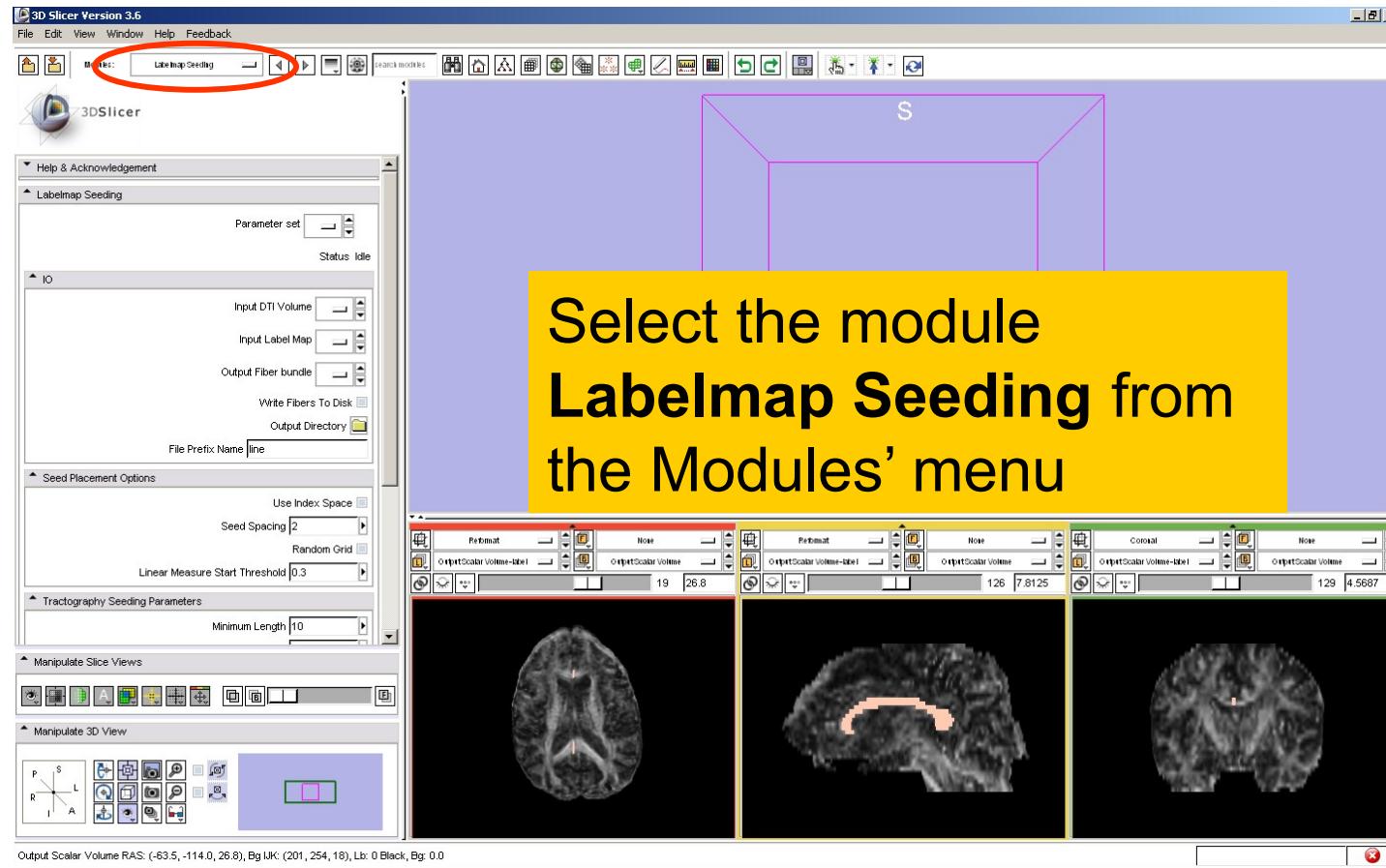
# LabelMap Generation

Select the label 2 (pink), click on the icon **Paint**, set the radius to **2** and draw a region of interest within the corpus callosum in the sagittal view on a set of 2 or 3 slices





# LabelMap Seeding





# LabelMap Seeding

Select the Input DTI volume  
**'Output DTI Volume'**

Select the Input Label Map  
**'Output Scalar Volume -label'**

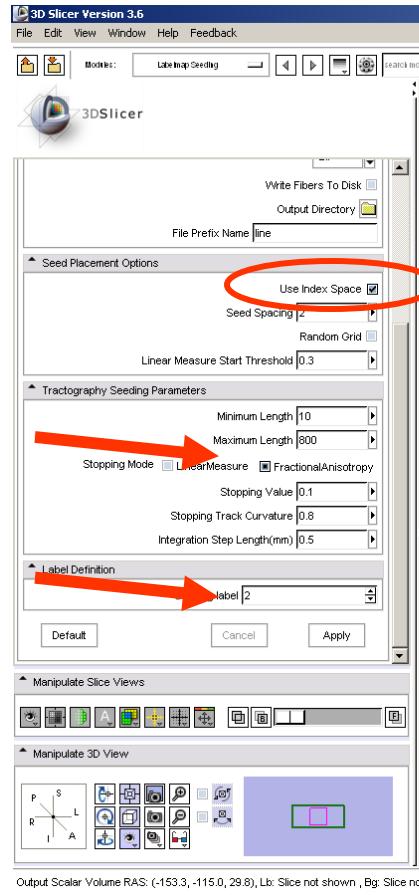
Select Output Fiber Bundle  
**'Create New Fiber Bundle'**

National Alliance for Medical Image Computing  
<http://na-mic.org> © 2010, ARR

Diffusion Tensor Imaging Tutorial, S.Pujol, PhD



# LabelMap Seeding



In the Seed Placement Options tab, select **Use Index Space**.

In the Tractography Seeding Parameters tab, select the ‘Stopping Mode’ **Fractional Anisotropy**, and use the default parameters for the minimum and maximum tract length, stopping value, stopping track curvature and integration step length.

In the Label Definition tab, set ‘Seeding label’ to label 2, and click on **Apply**

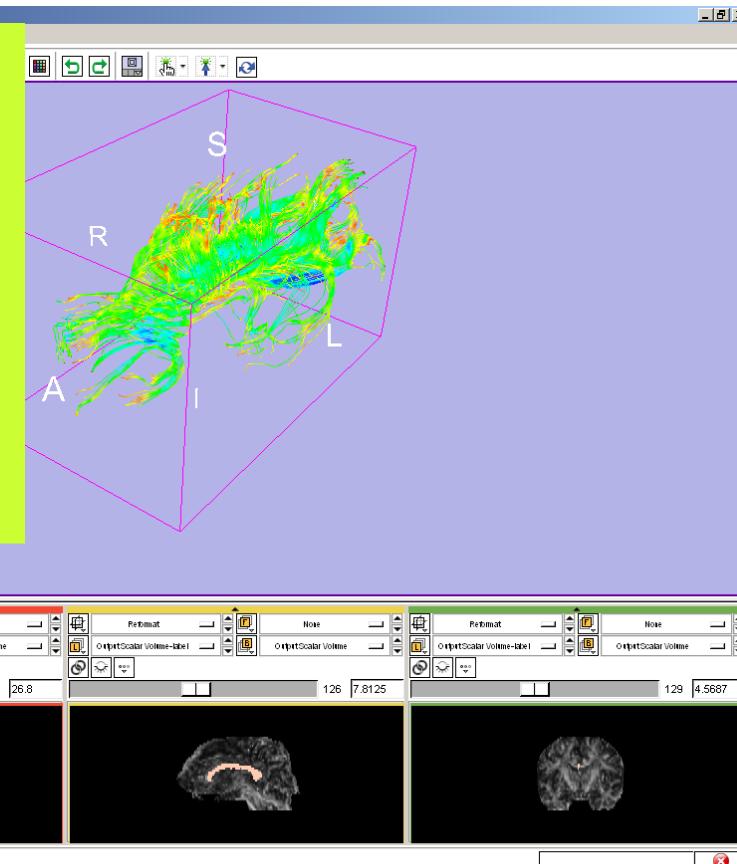


# LabelMap Seeding

3D Slicer Version 3.6

The tracts generated within the corpus callosum region appear in the 3DViewer.

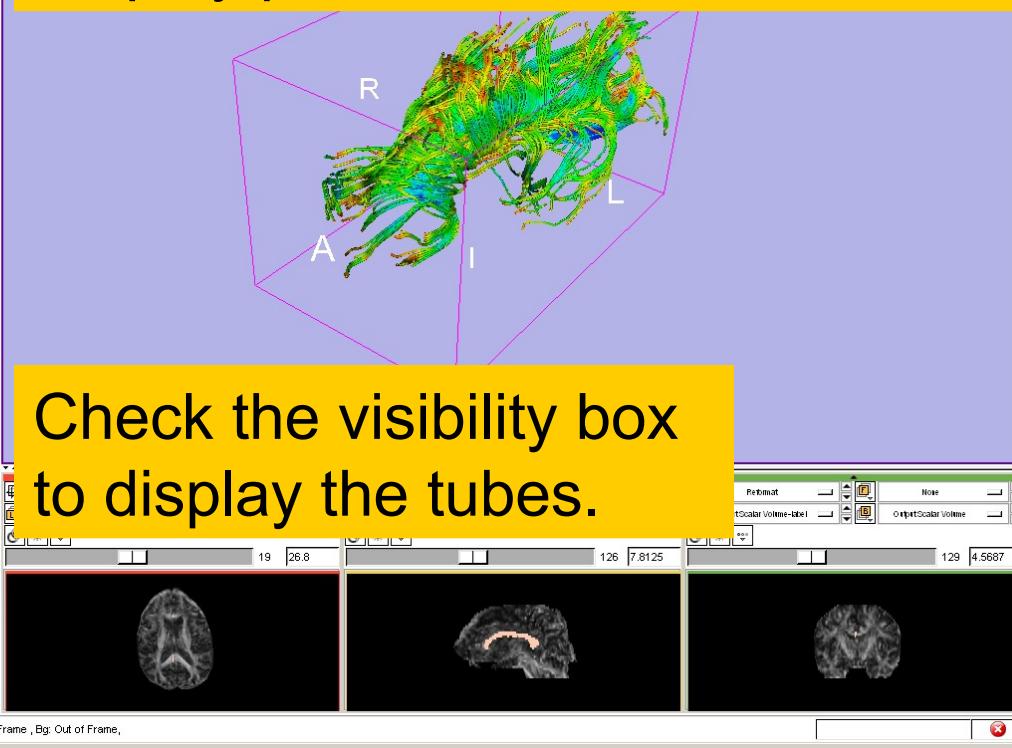
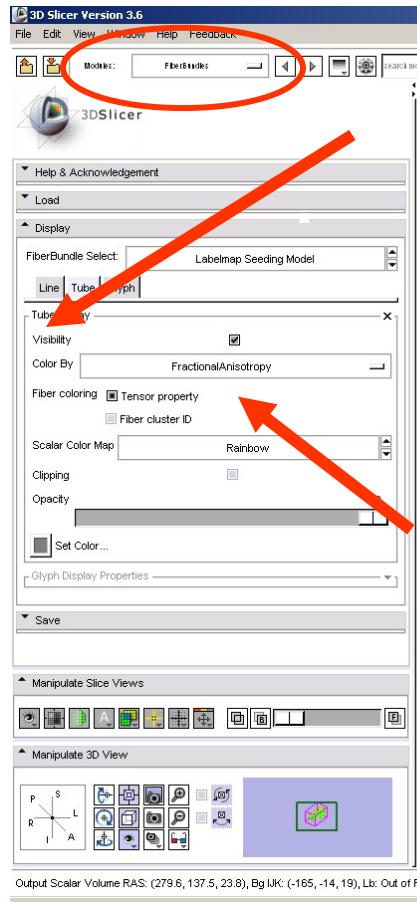
The color map used represent the FA values along the tracts.





# LabelMap Seeding

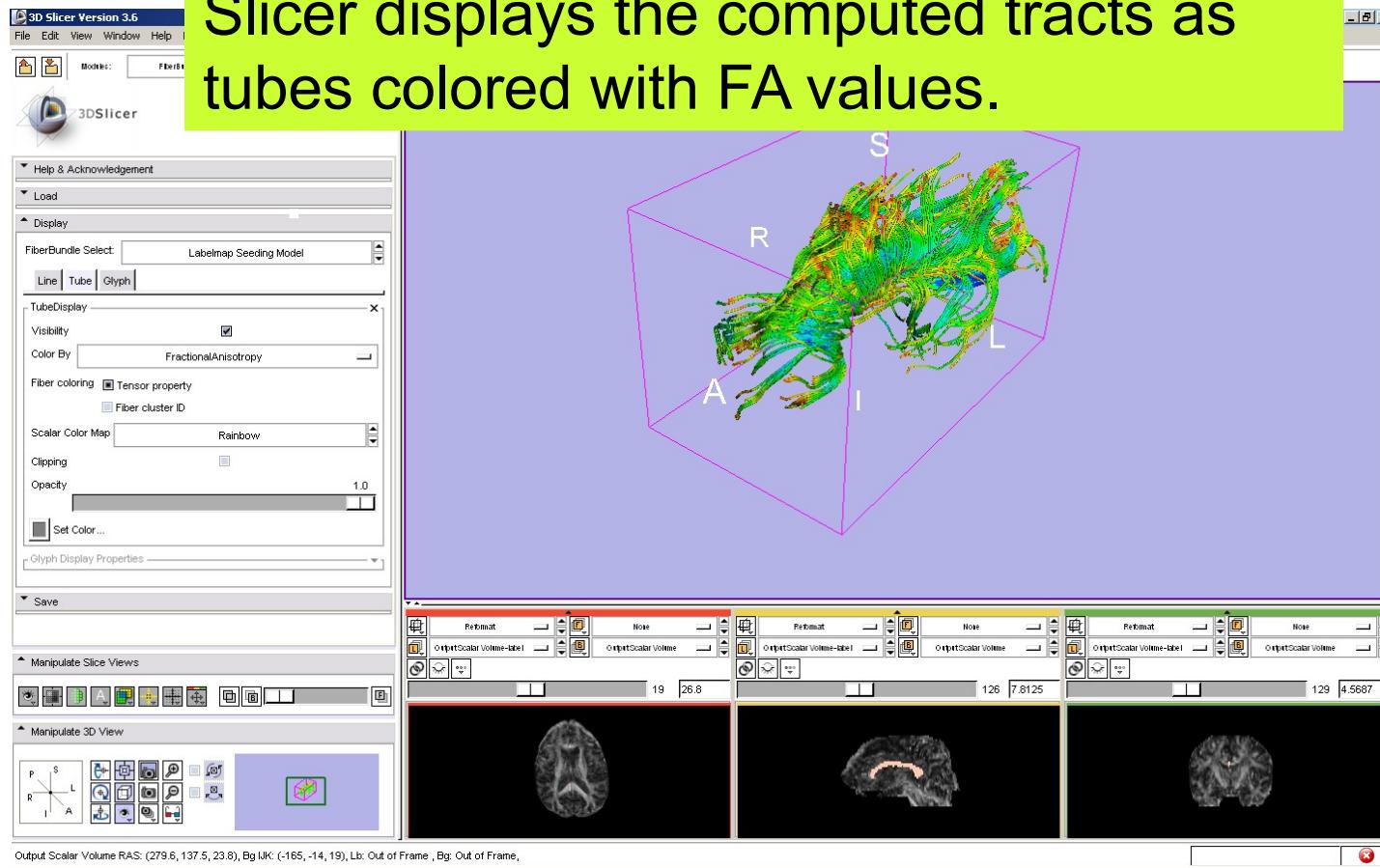
Select the module **FiberBundles**, and click on the tab **Tube** in the Display panel





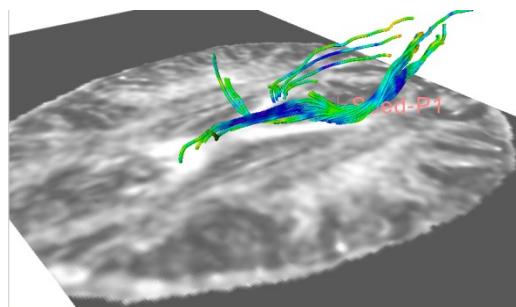
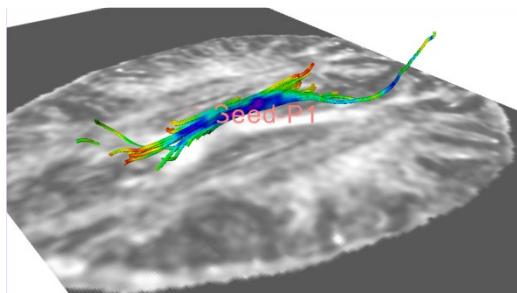
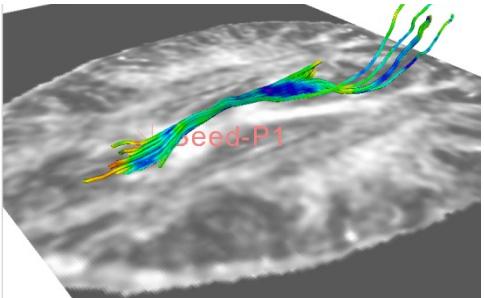
# LabelMap Seeding

Slicer displays the computed tracts as tubes colored with FA values.



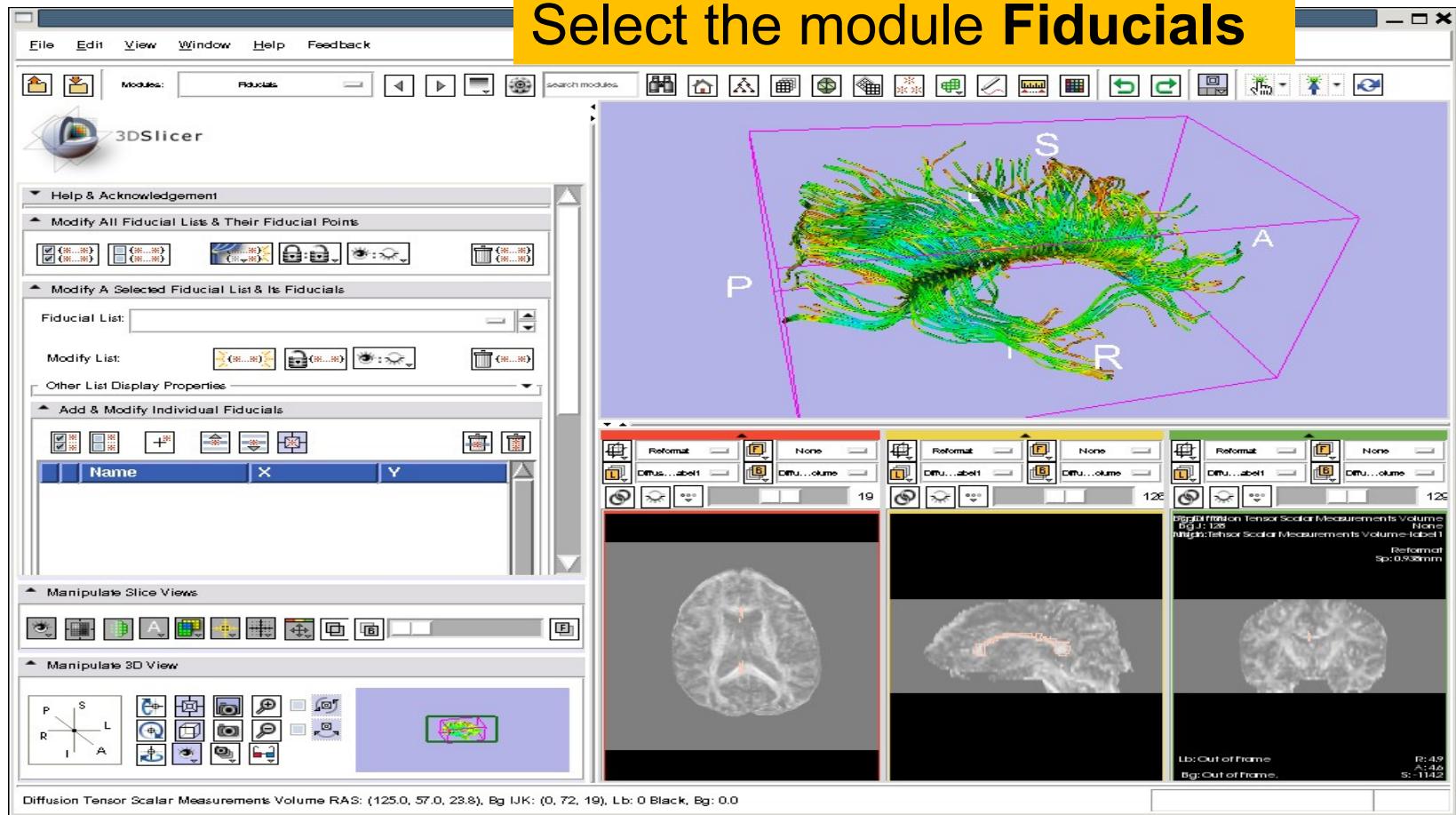
## Part 4:

# Tractography on-the-fly



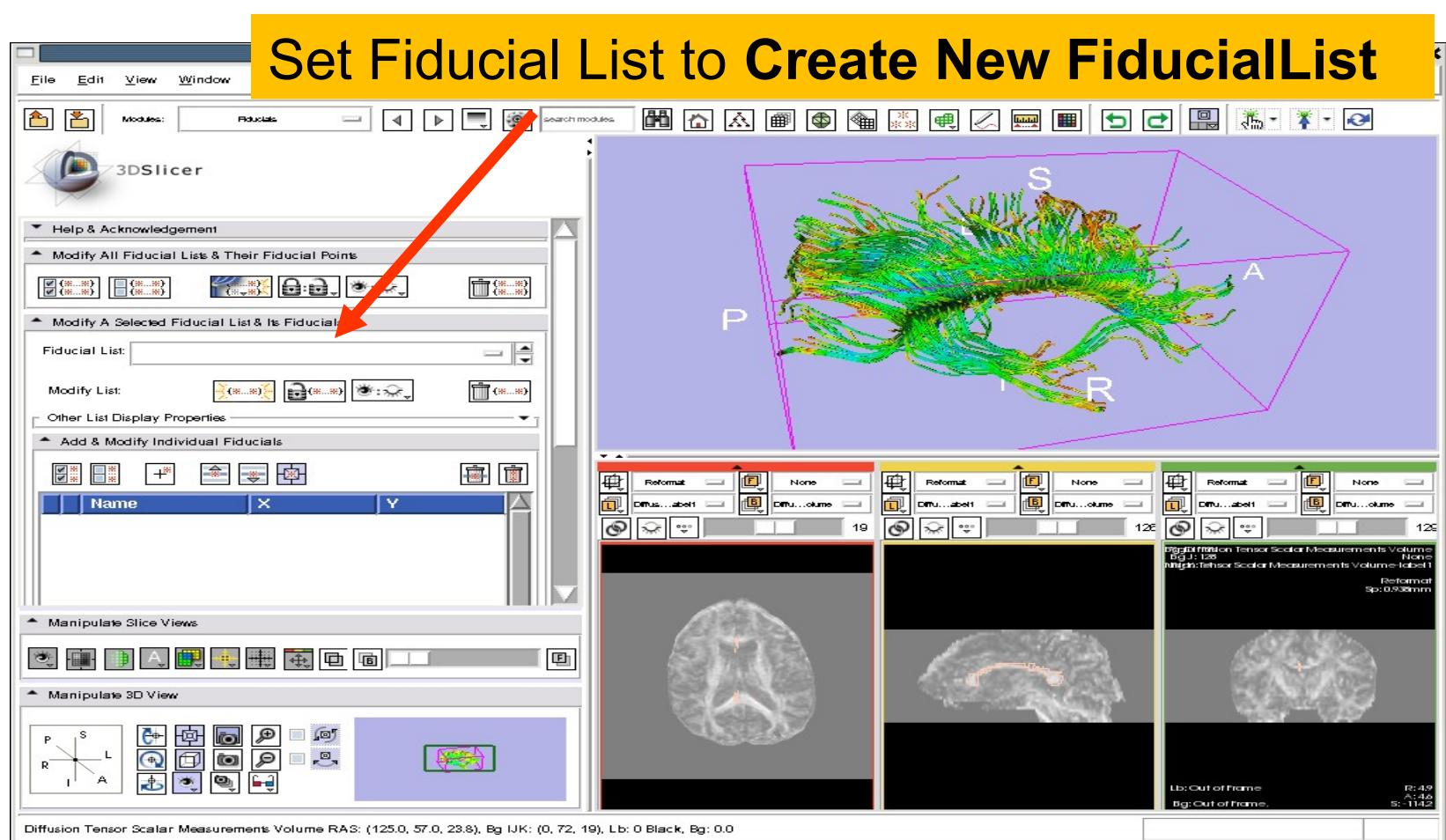


# Fiducial Seeding



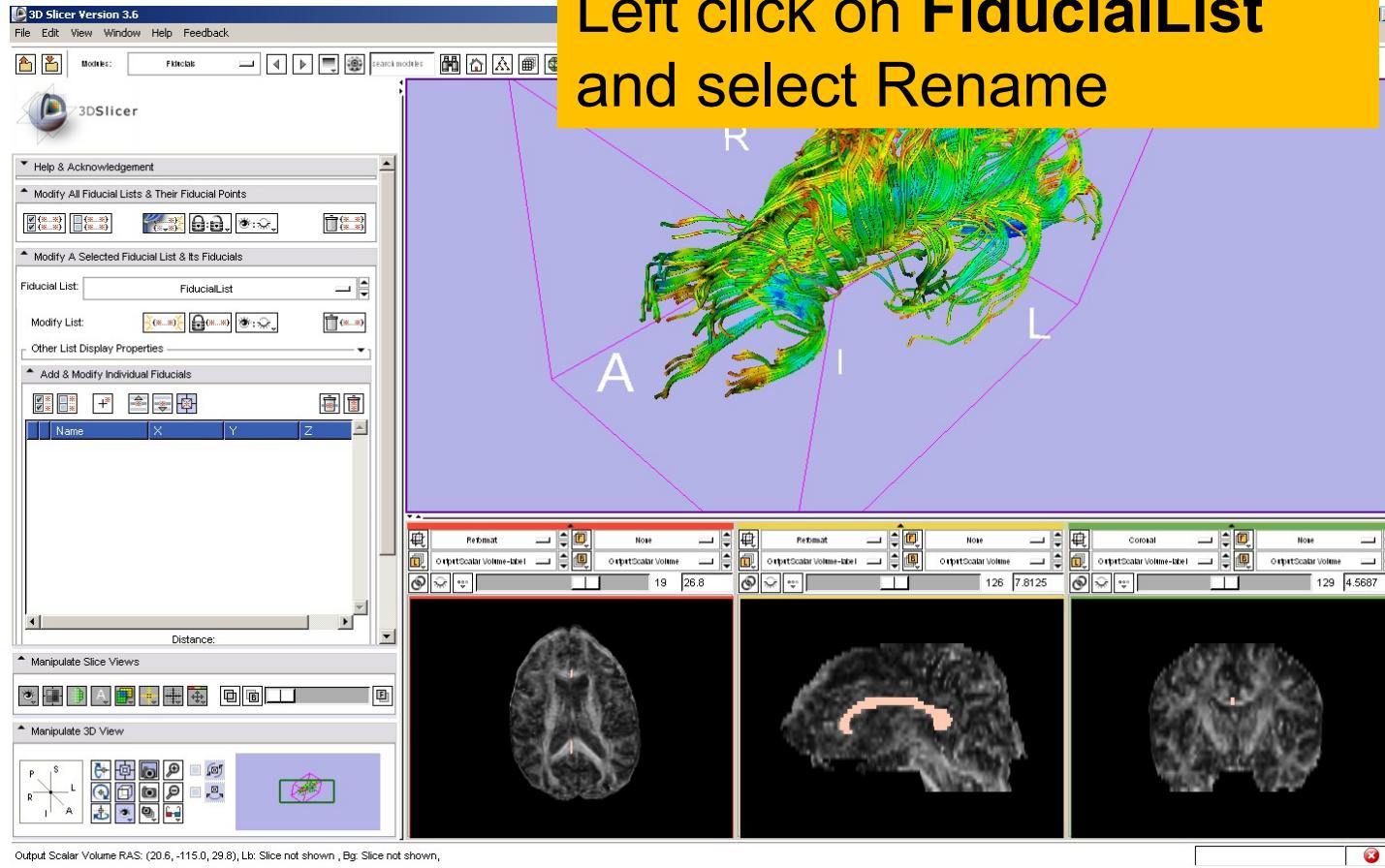


# Fiducial Seeding





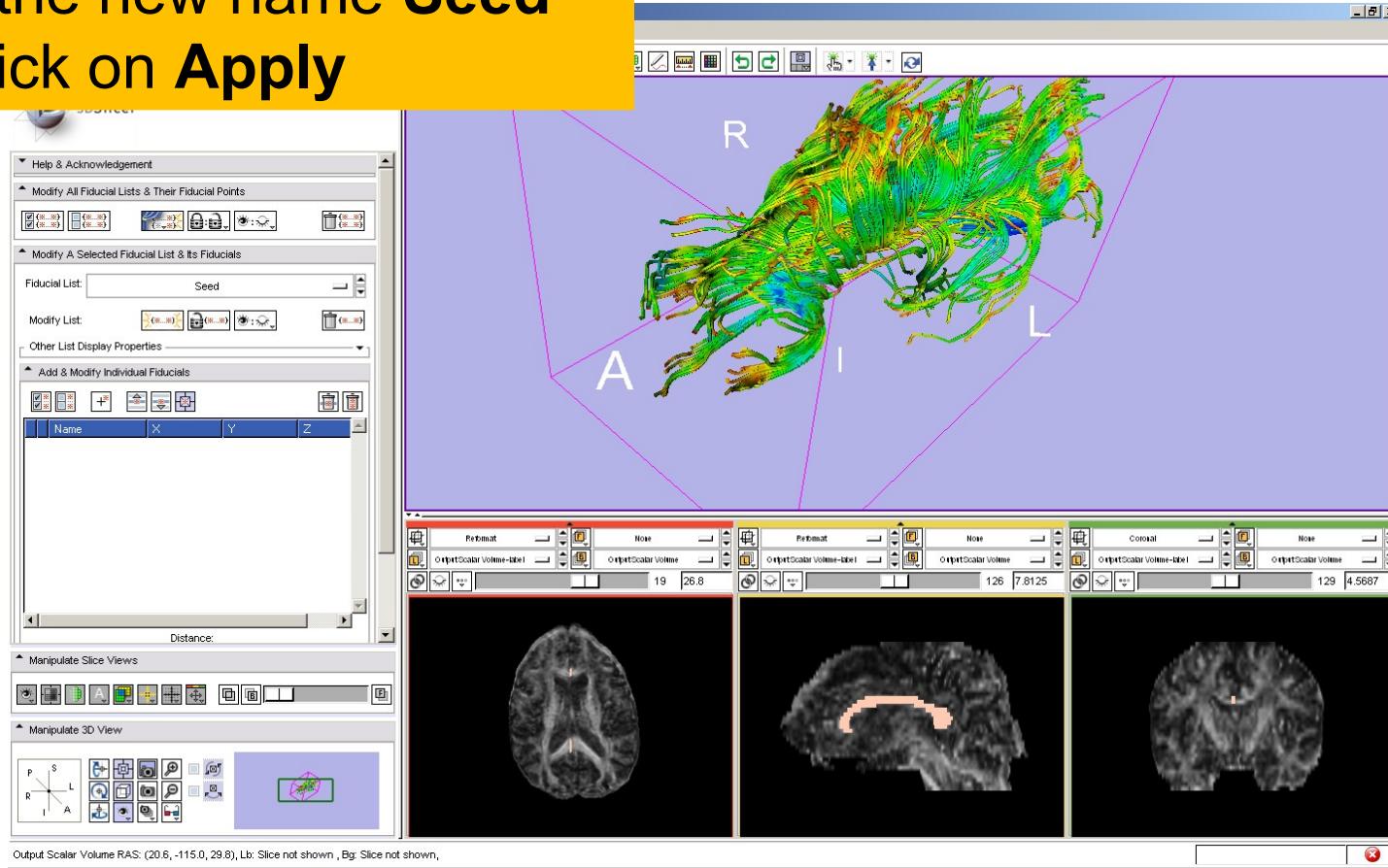
# Fiducial Seeding





# Fiducial Seeding

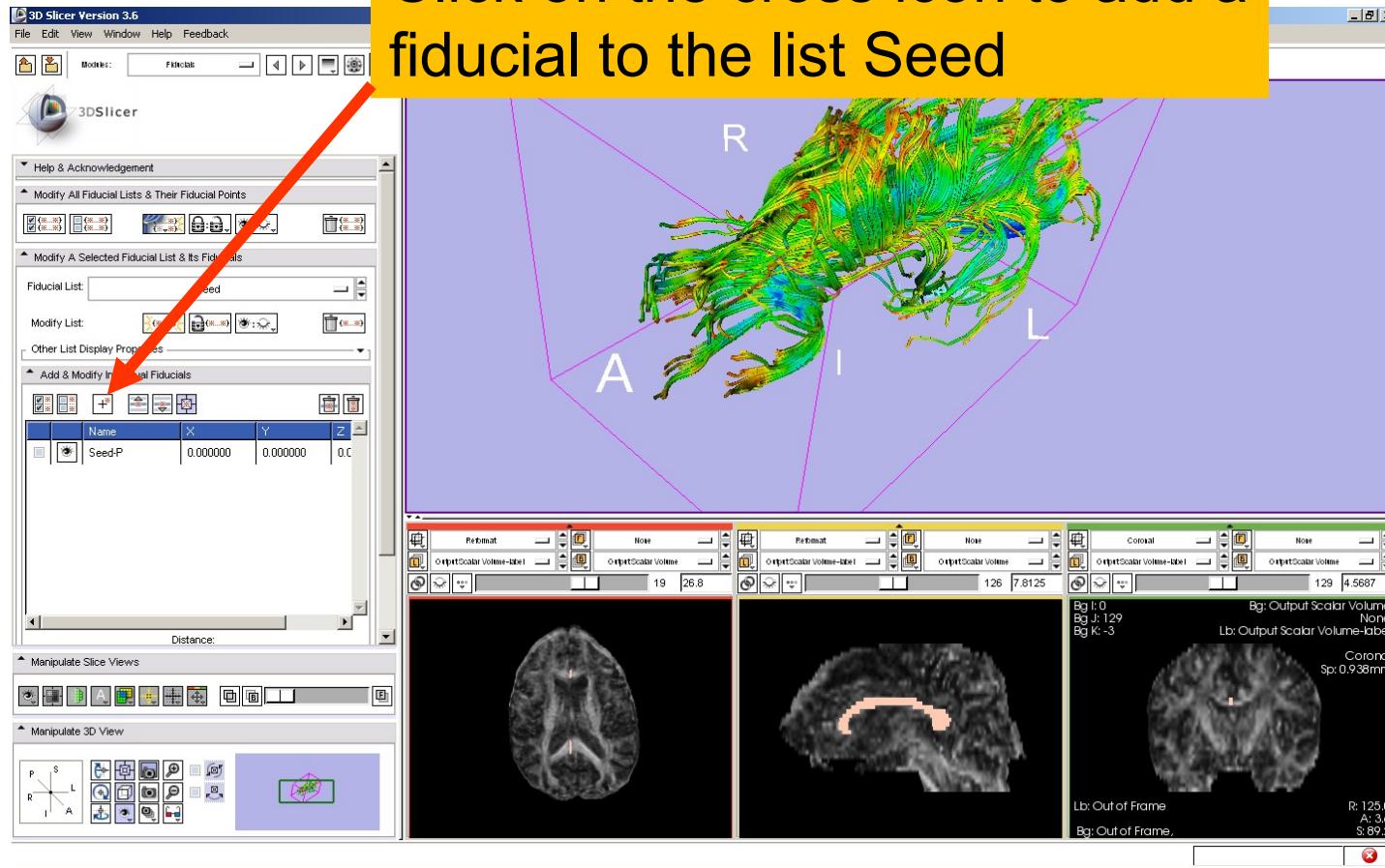
Enter the new name **Seed**  
and click on **Apply**





# Fiducial Seeding

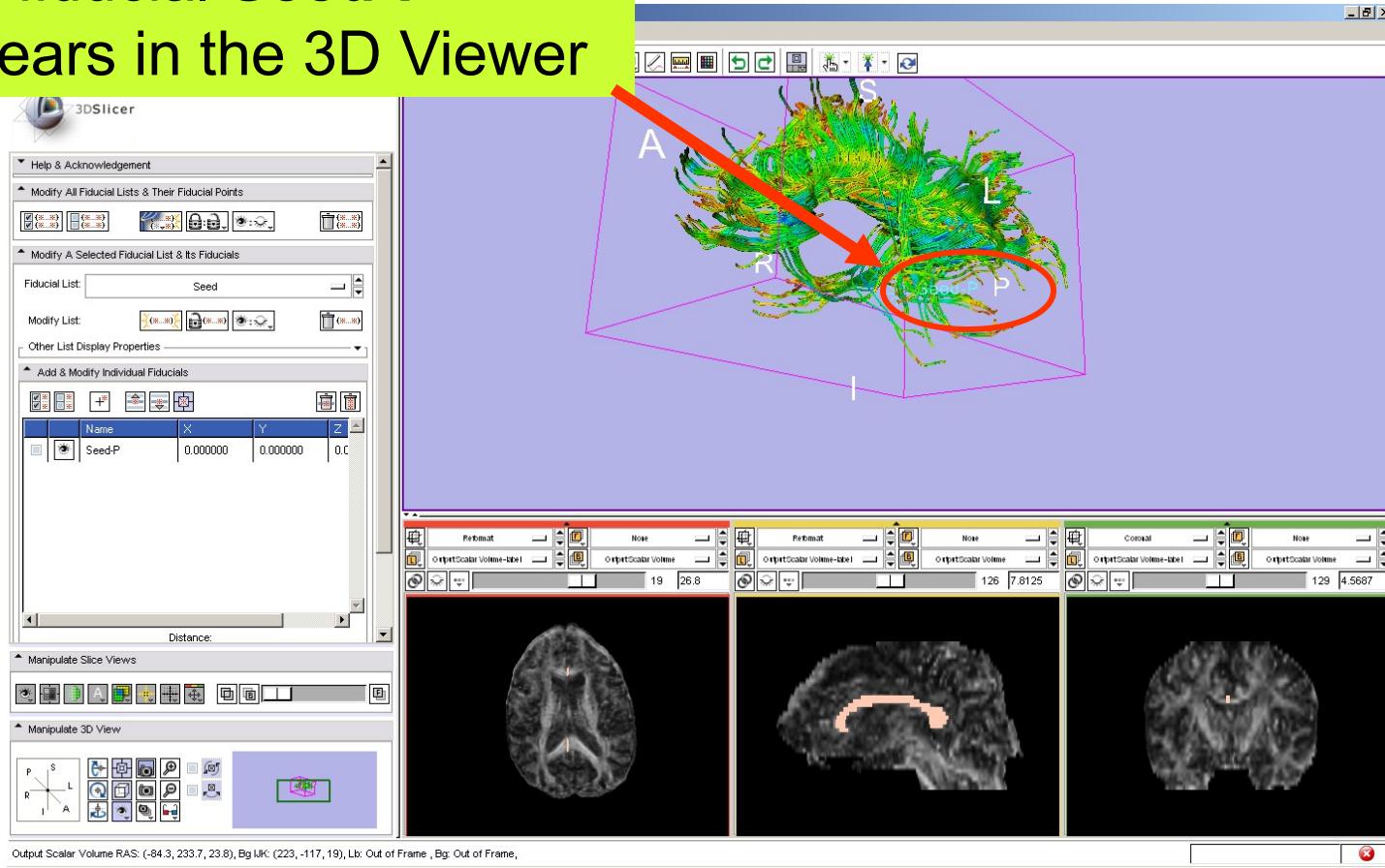
Click on the cross icon to add a fiducial to the list Seed





# Fiducial Seeding

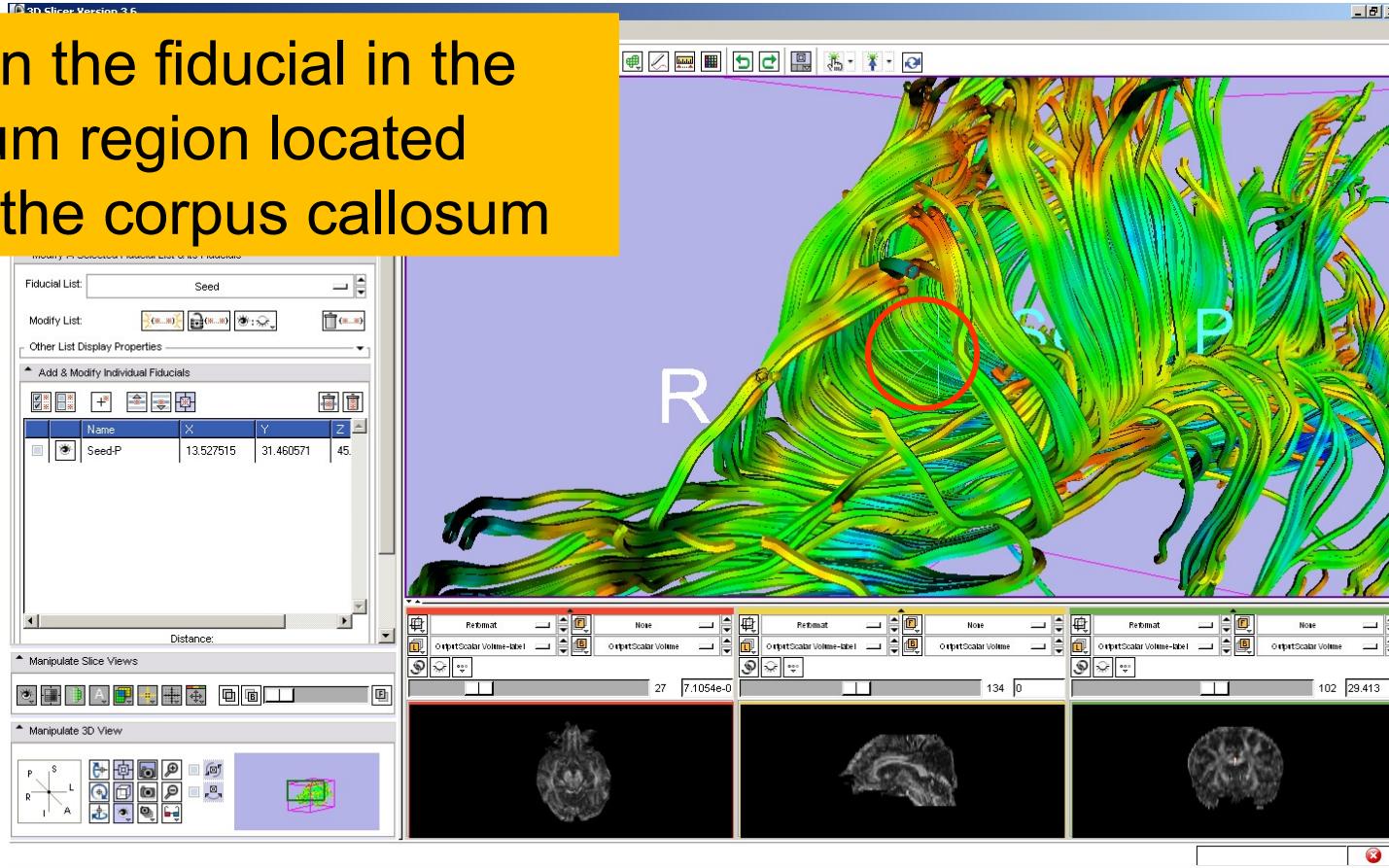
The fiducial Seed-P appears in the 3D Viewer





# Fiducial Seeding

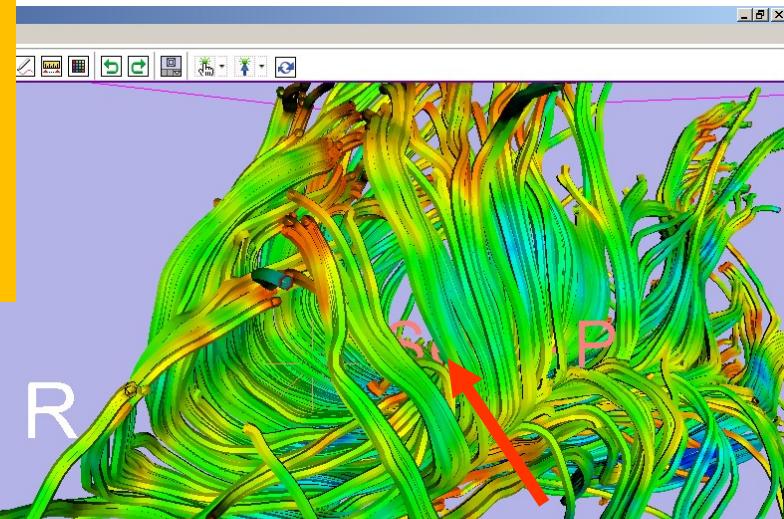
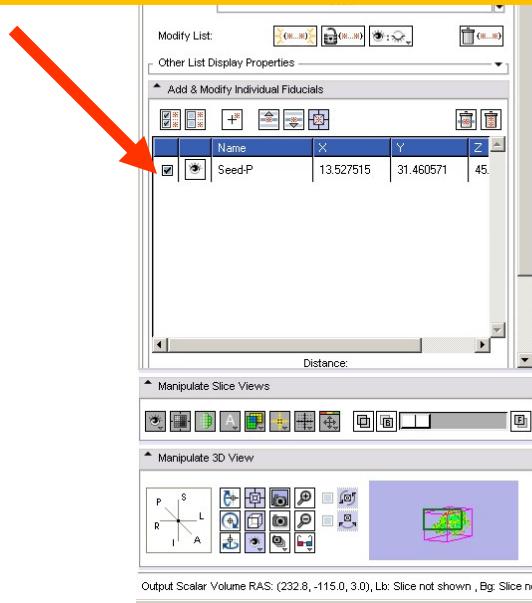
Position the fiducial in the cingulum region located above the corpus callosum





# Fiducial Seeding

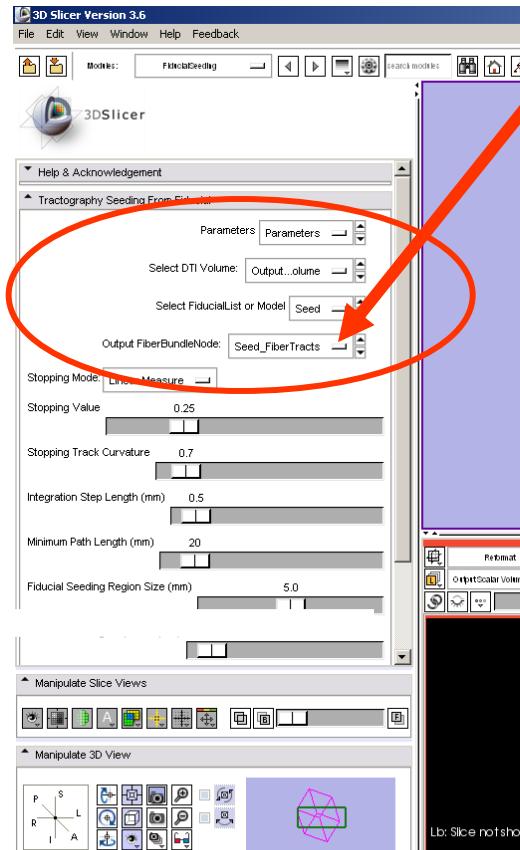
**Check the box to select the ‘Seed-P’ fiducial:** we will use this fiducial to drive the tractography



Once selected, the fiducial Seed-P is displayed in pink letters in the 3D viewer.



# Fiducial Seeding



**Set the Output FiberBundleNode to Create New FiberBundle**

**Important:** this step **must** be done first

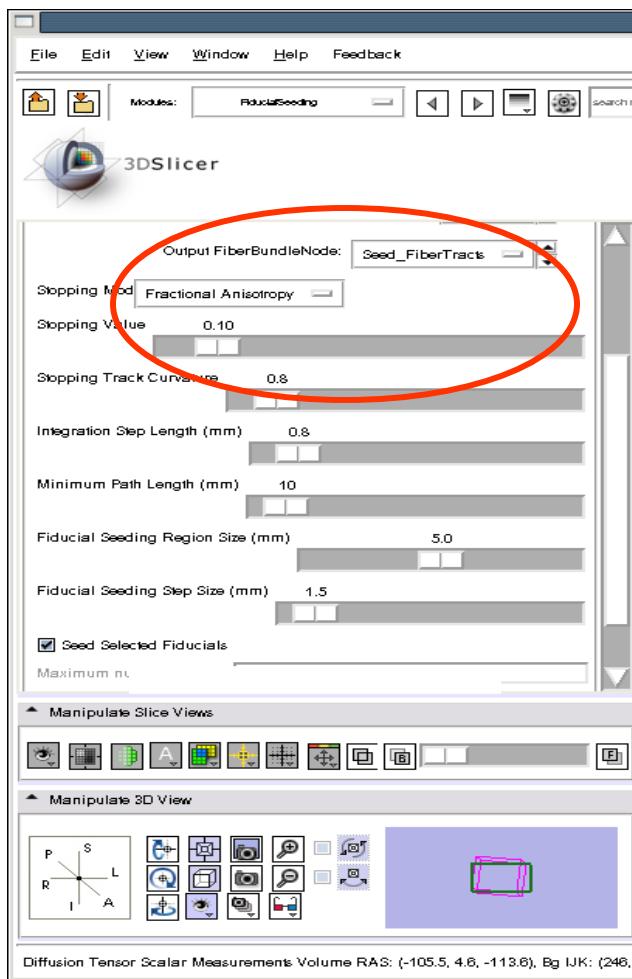
**Select the module Fiducial Seeding**

**Set the DTI Volume to Output DTI Volume**

**Select the Fiducial List Seed**



# Fiducial Seeding



Set the Stopping Mode to **Fractional Anisotropy** and set the tractography parameters to the values that we used for the corpus callosum:

**Stopping Value: 0.1**

**Stopping Track Curvature: 0.8**

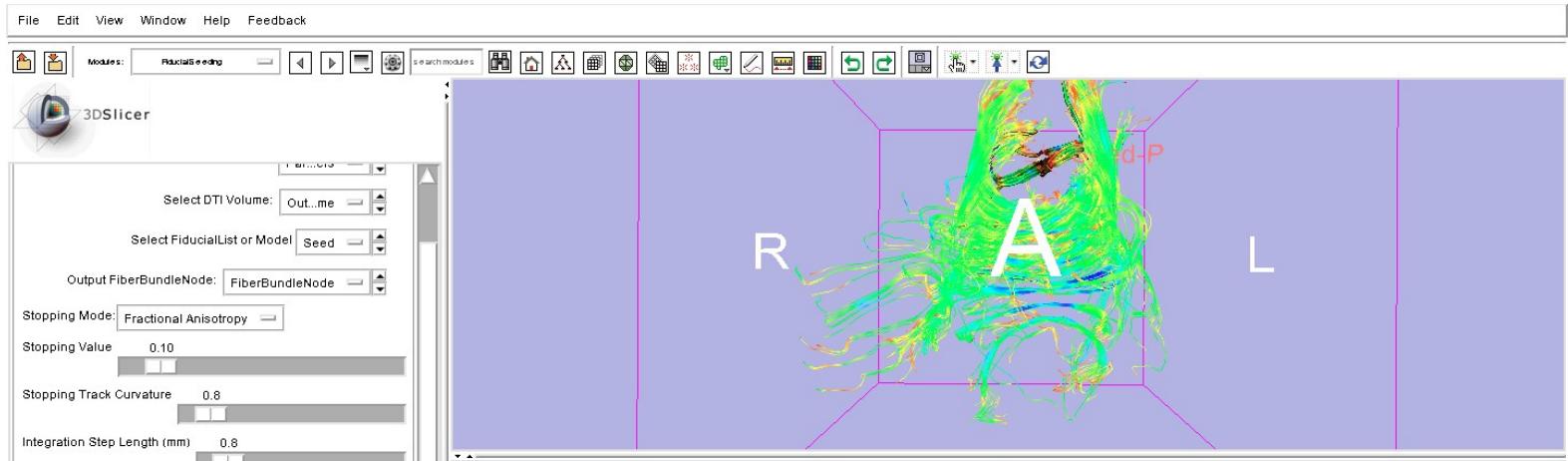
**Step Length: 0.8 mm**

**Minimum Path Length: 10 mm**

**Fiducial Stepping Size: 1.5 mm**



# Fiducial Seeding



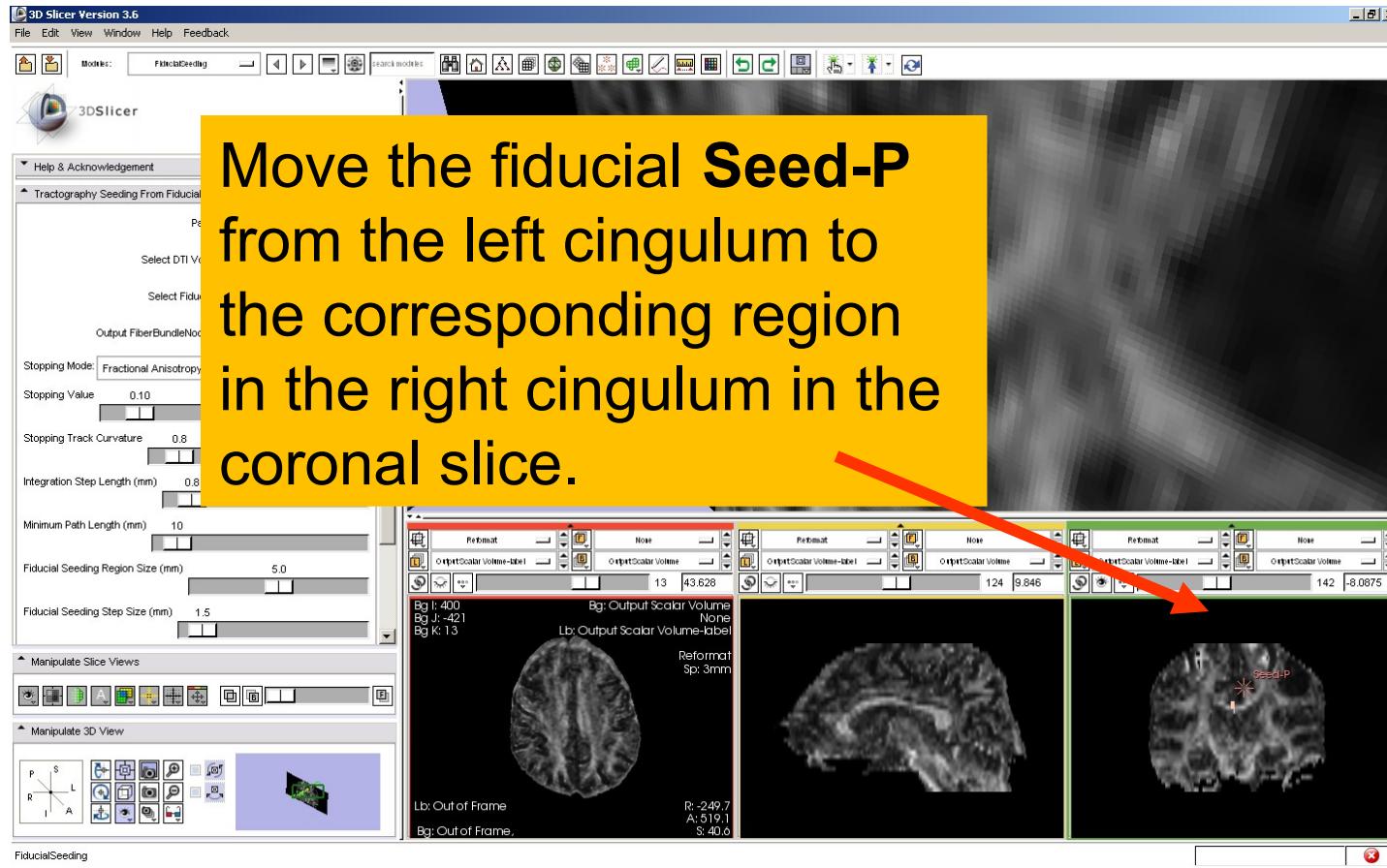
Slicer displays the tracts seeded from the Fiducial Seed-P.

The tracts correspond to the region of the cingulum located above the corpus callosum.

For better visualization, uncheck the visibility box under **Tubes** in the **Fiber Bundles** module (Slide 42).

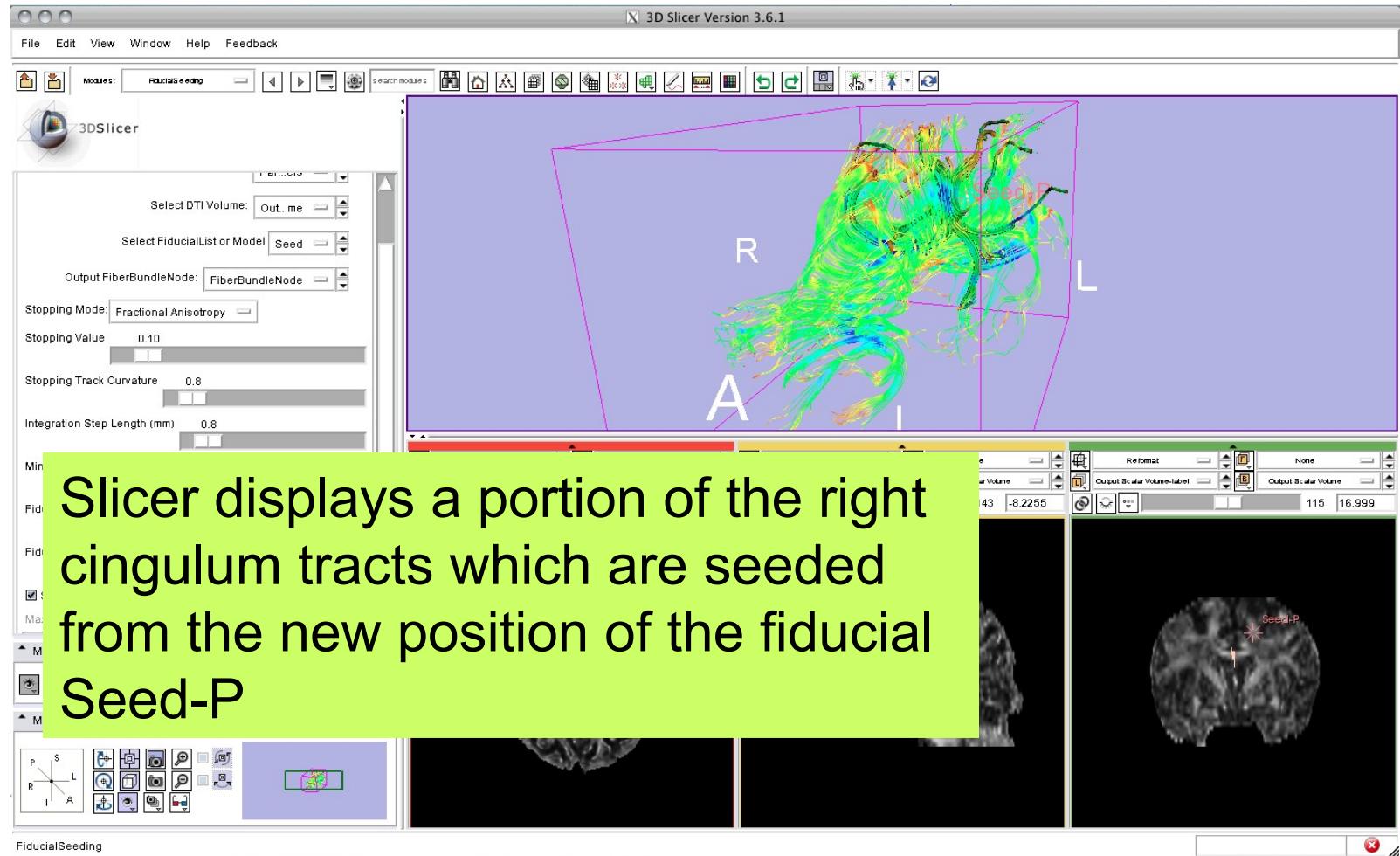


# Fiducial Seeding



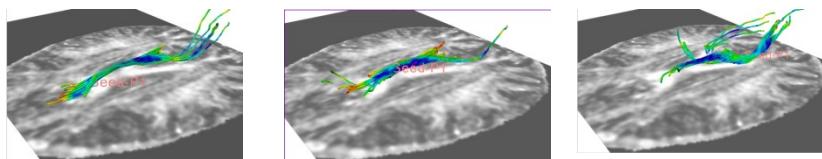
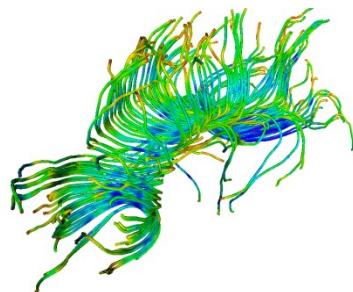
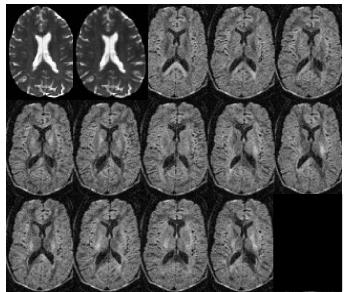


# Fiducial Seeding



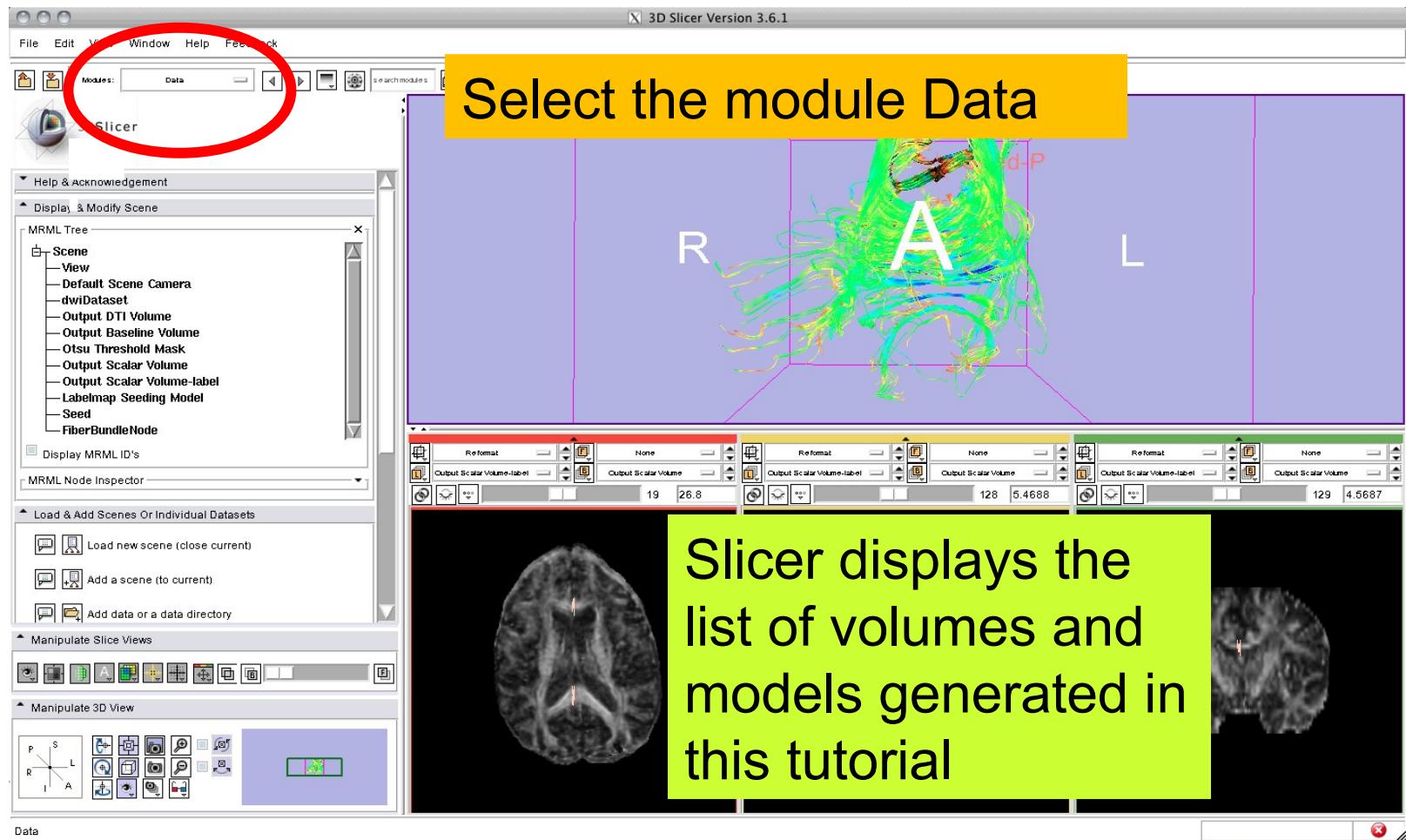
# Part 5:

# Saving a DTI Scene



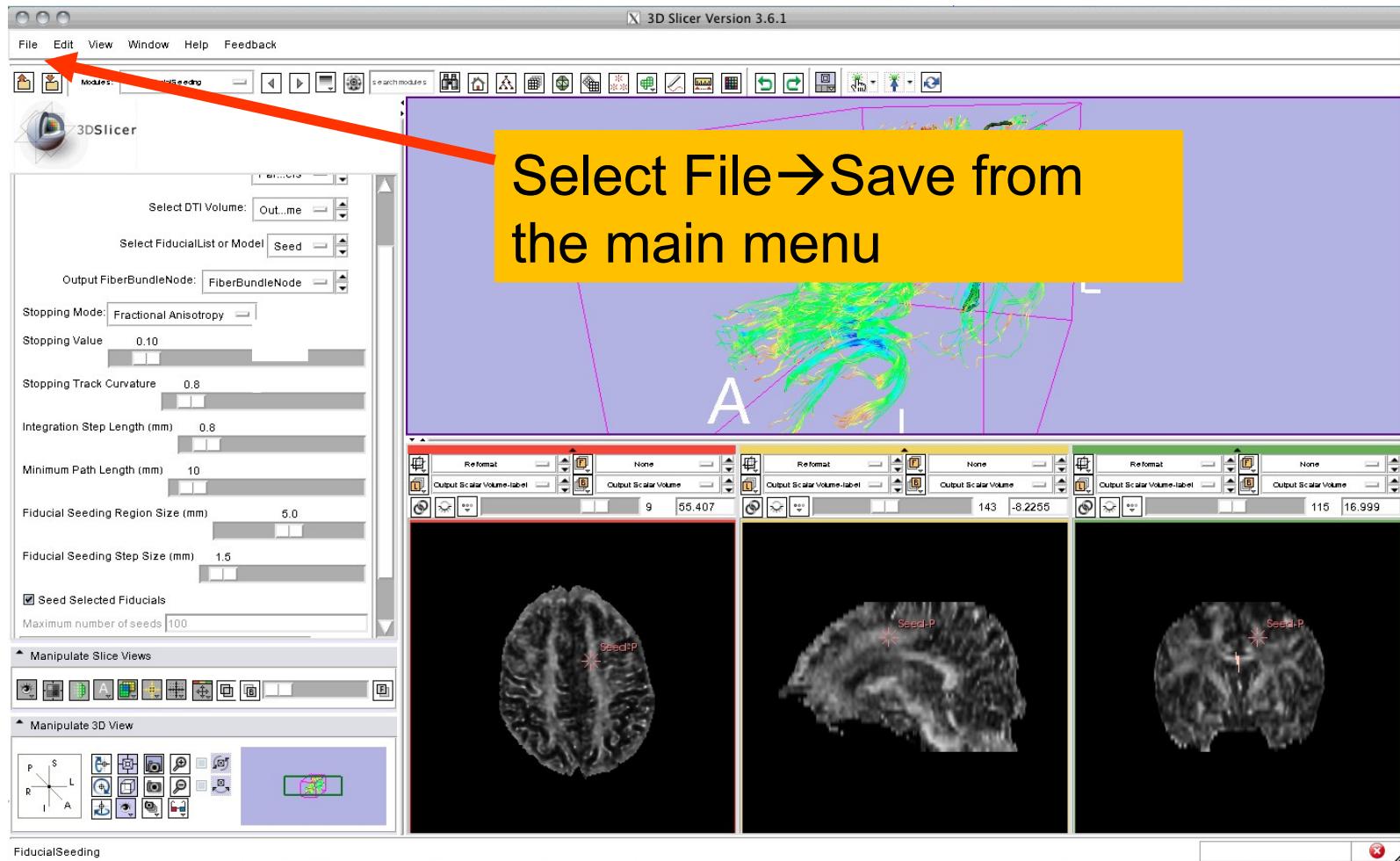


# DTI Scene





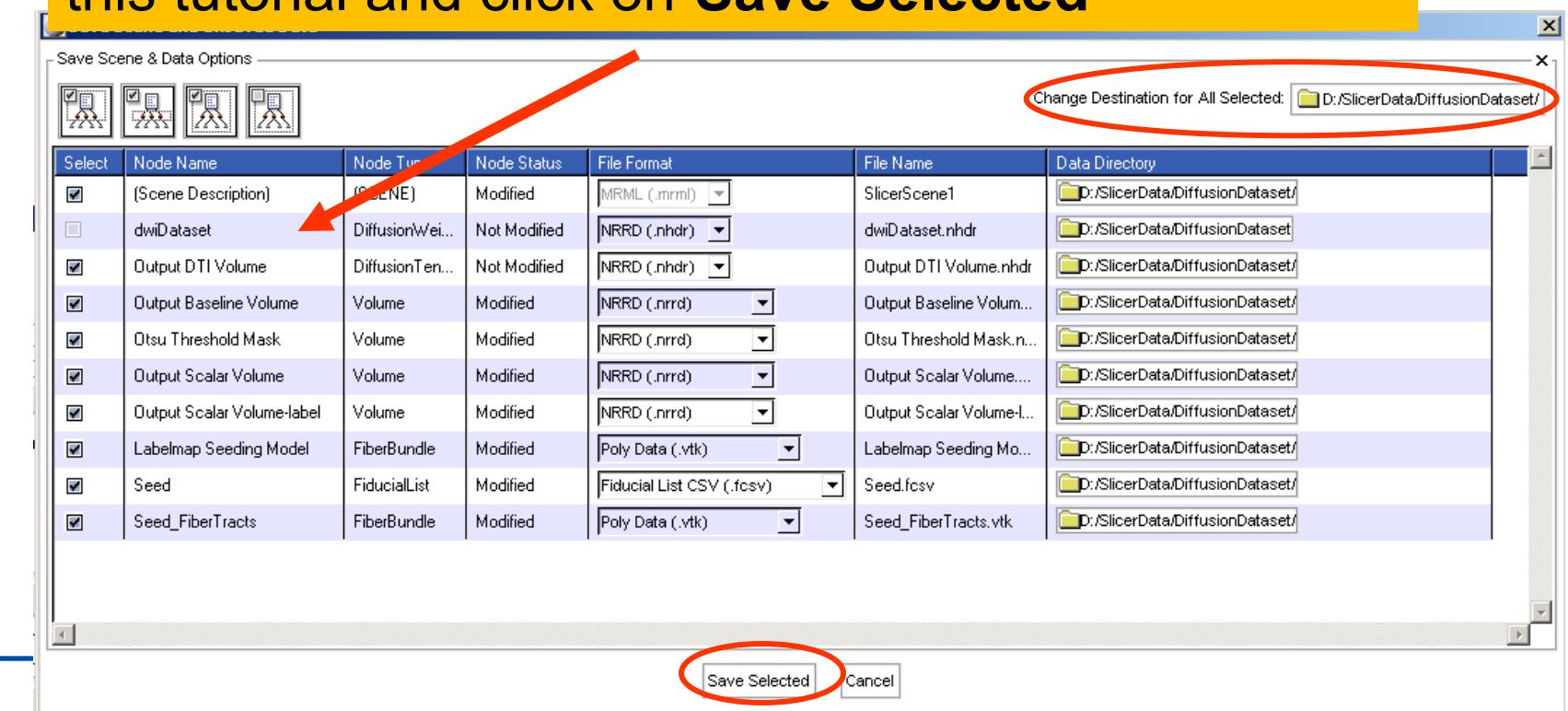
# Saving a DTI Scene





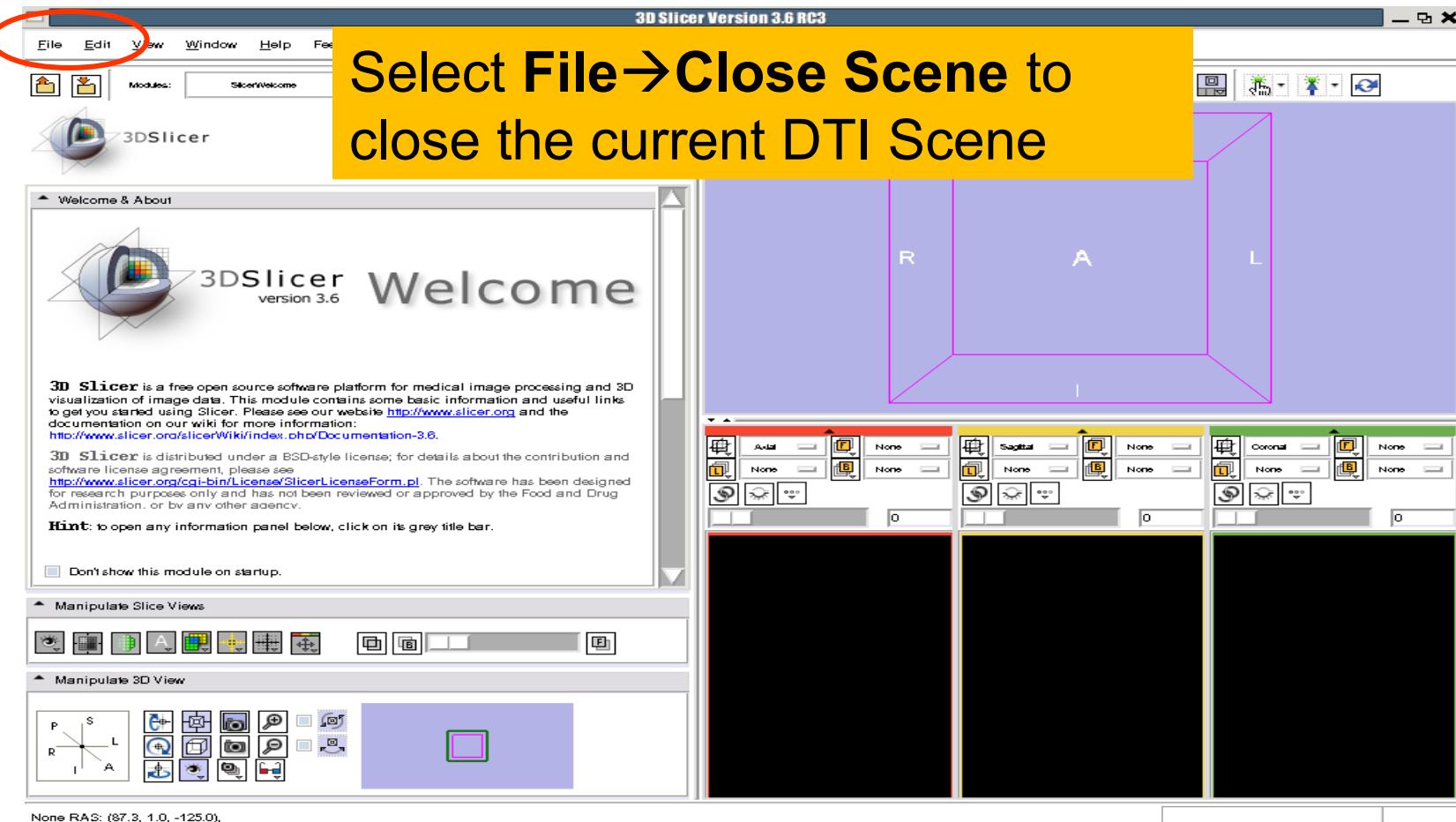
# Saving a DTI Scene

Browse to a directory where you would like to save the data. Once you have selected a directory, select all the files that have been created during this tutorial and click on **Save Selected**



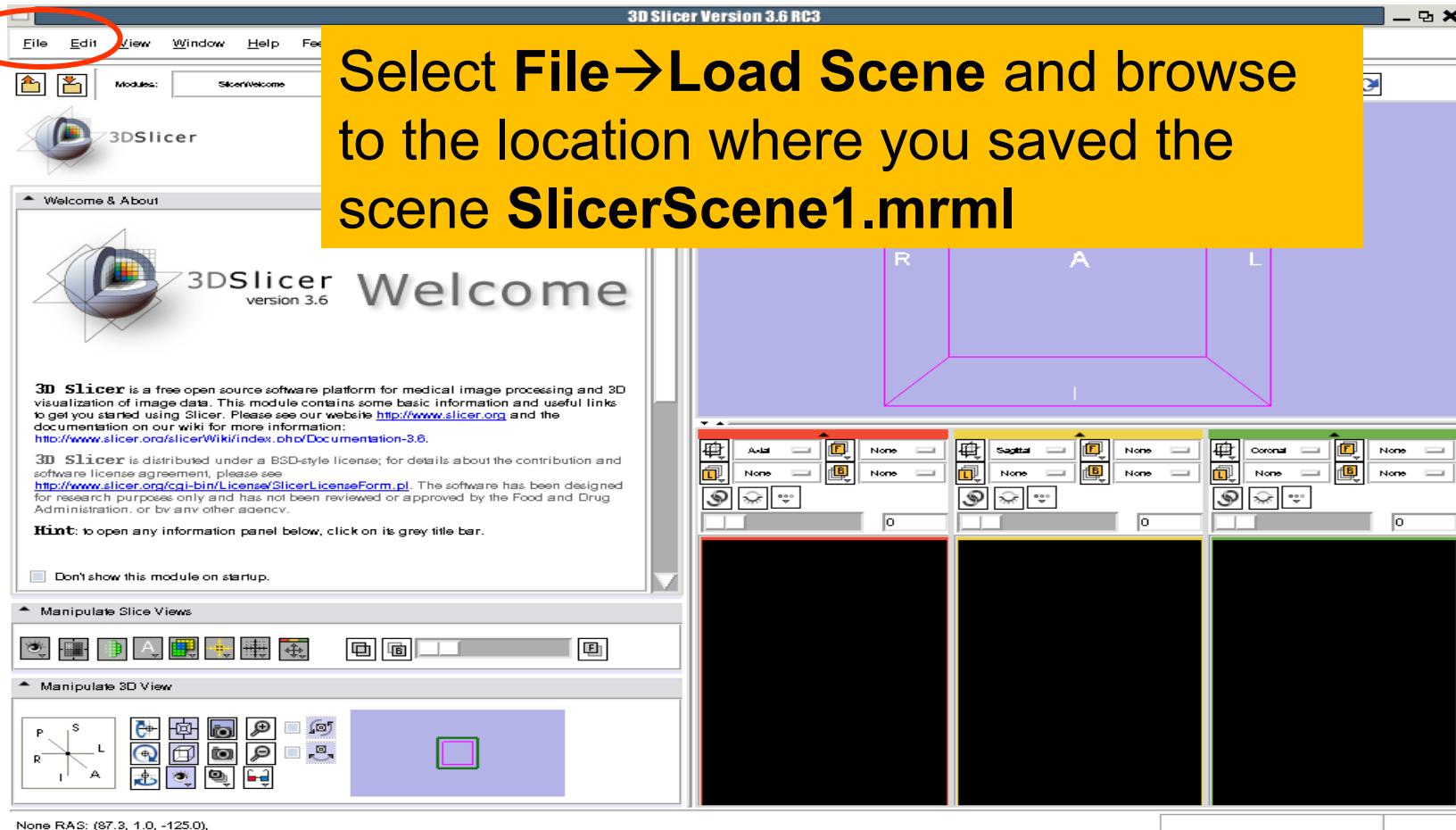


# Saving a DTI Scene



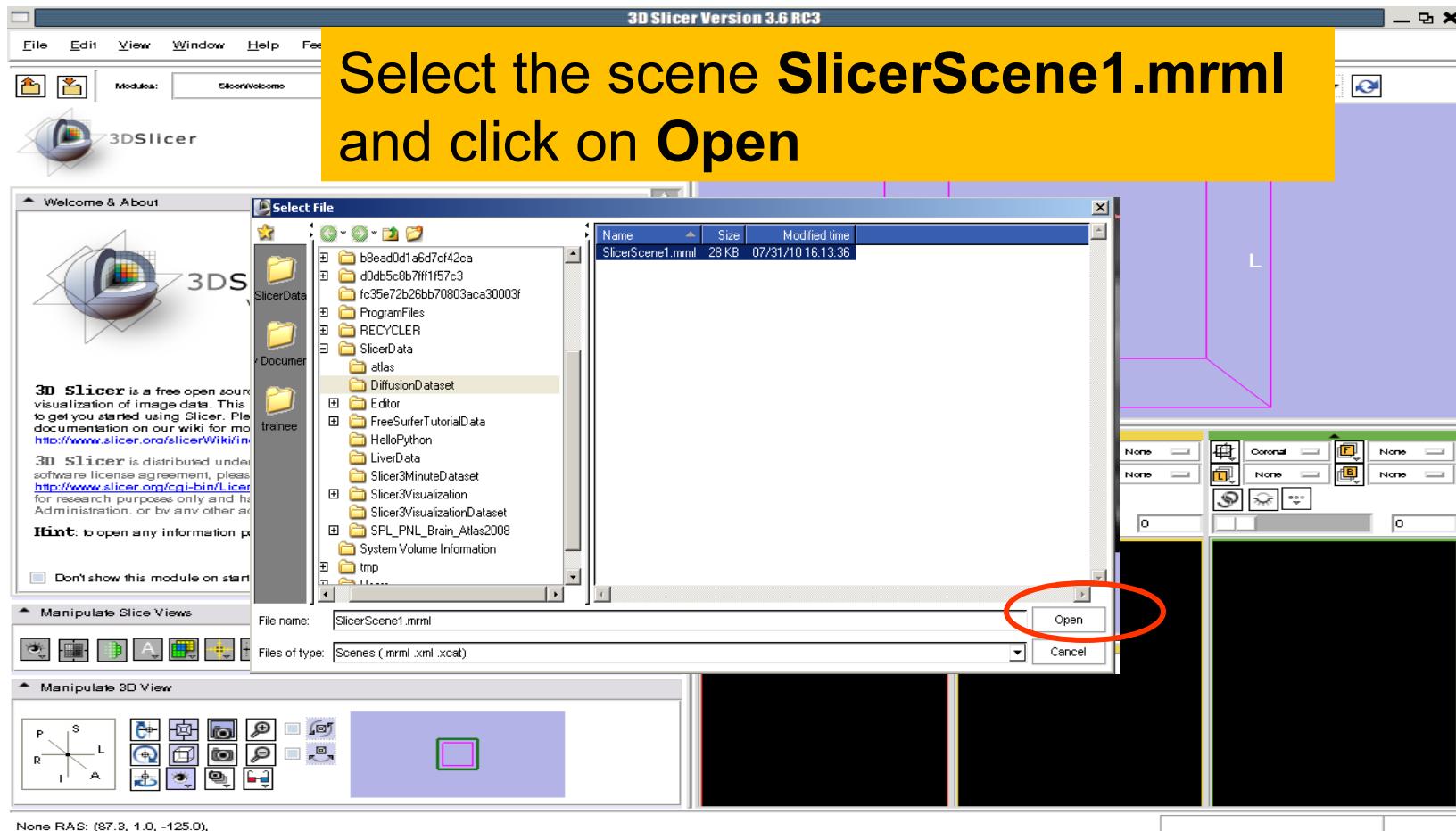


# Loading a DTI Scene



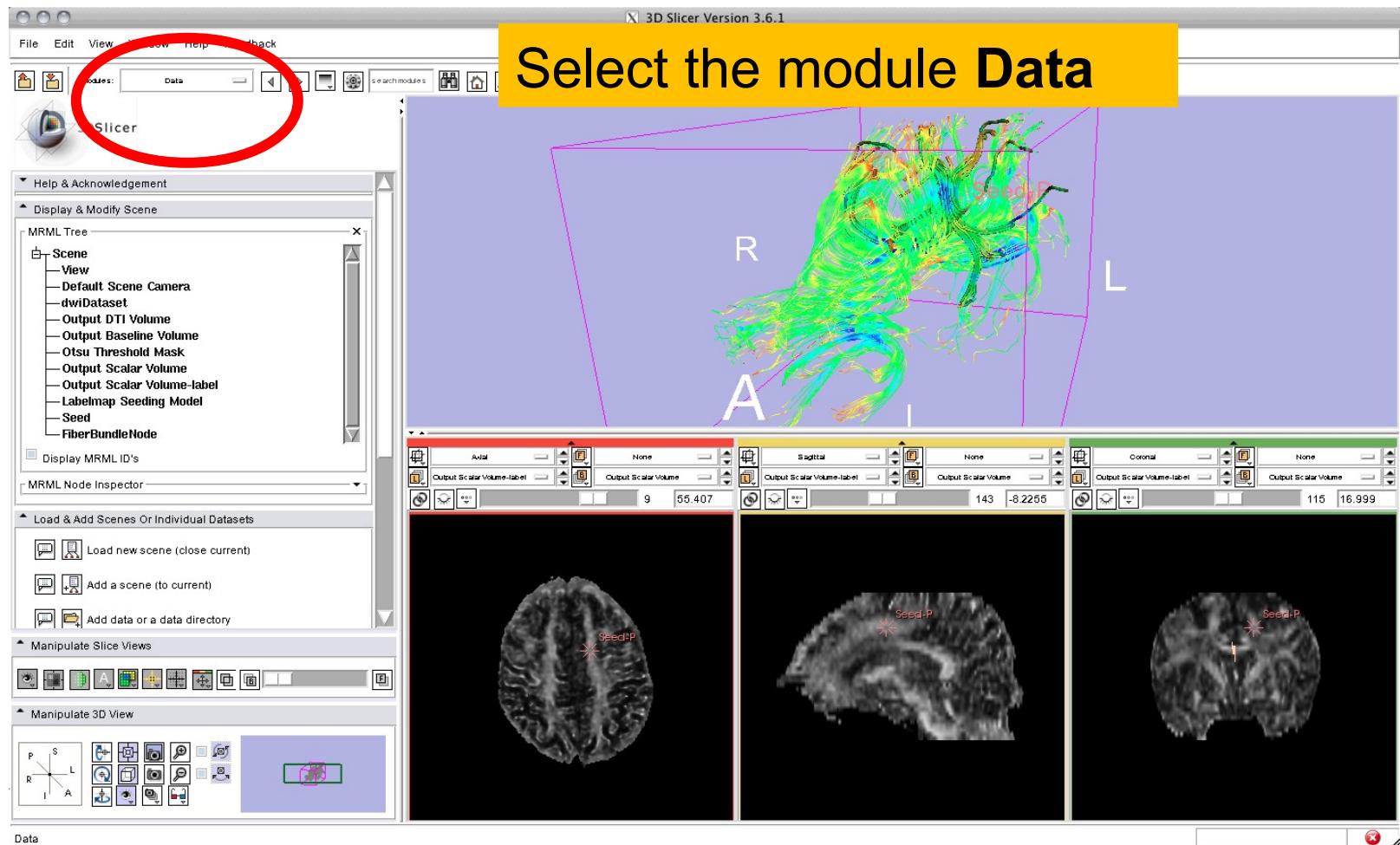


# Loading a DTI Scene



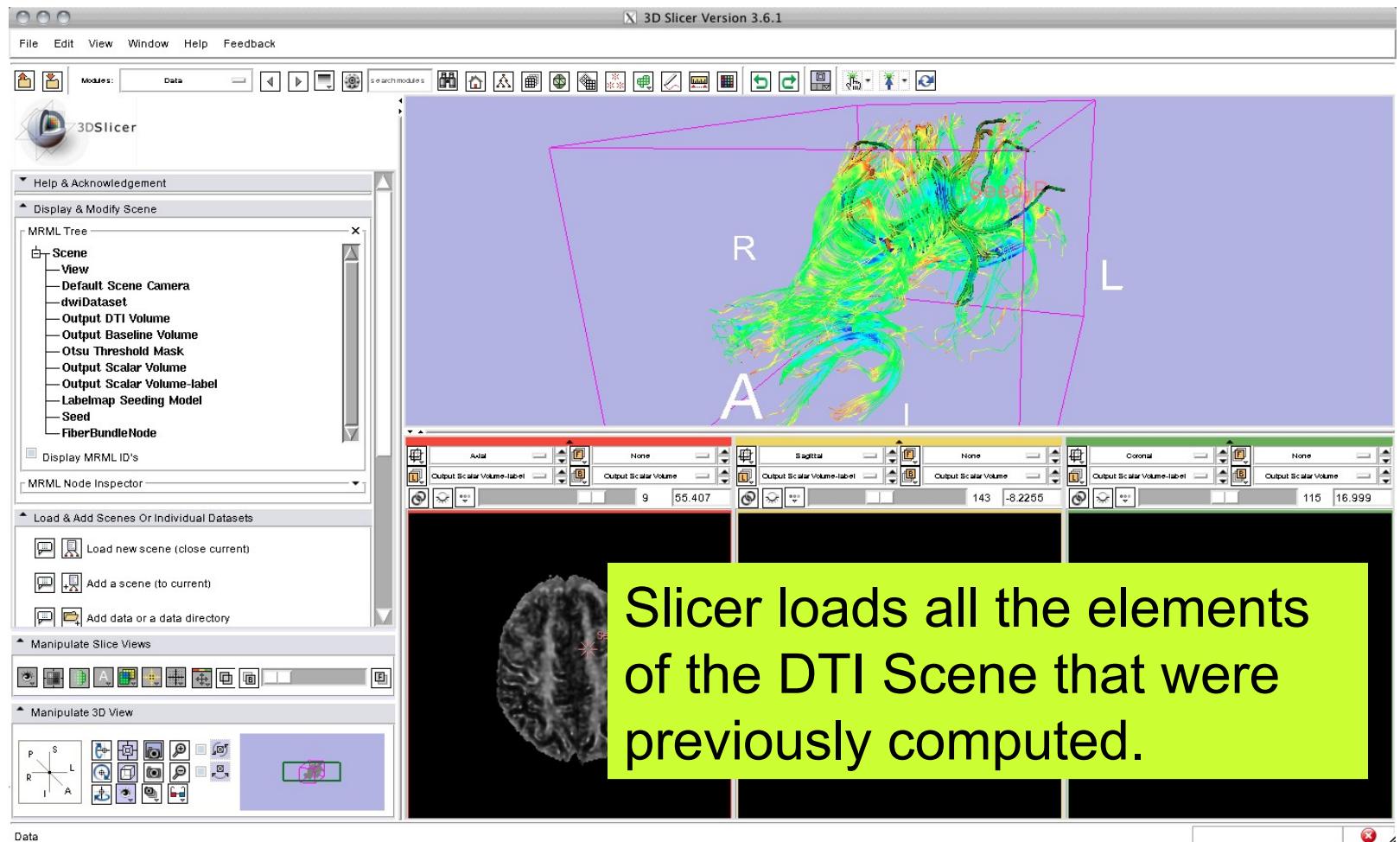


# Loading a DTI Scene





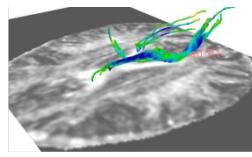
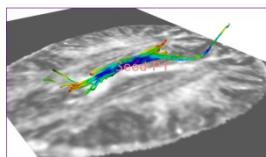
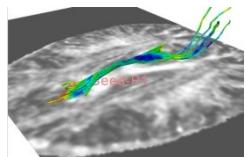
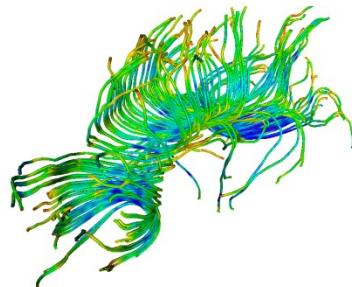
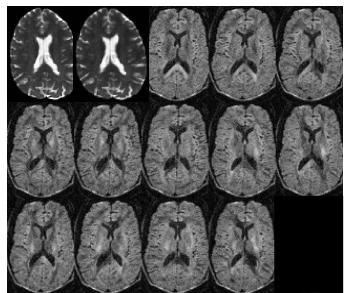
# Loading a DTI Scene



Slicer loads all the elements of the DTI Scene that were previously computed.



# Conclusion



This tutorial guided you through some of the **Diffusion MR** capabilities of the **Slicer3** software for studying the brain white matter pathways.

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# Acknowledgments

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